

Brazos River Authority FY 2026– 2027 Clean Rivers Program Quality Assurance Project Plan

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**Clean Rivers Program
Water Quality Planning Division
Texas Commission on Environmental Quality
P.O. Box 13087, MC 234
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Effective Period: FY 2026 to FY 2027**

Questions concerning this QAPP should be directed to:

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A2 Approval Page

Texas Commission on Environmental Quality

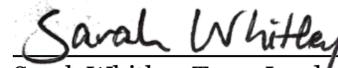
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Water Quality Monitoring and Assessment
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8/20/2025

Date



Sarah Whitley, Team Leader
Water Quality Standards and Clean Rivers Program

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Sunshyne Hendrix
Project Quality Assurance Specialist
Clean Rivers Program

8/20/2025

Date



Sunshyne Hendrix, Project Manager
Clean Rivers Program

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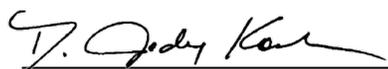


Cathy Anderson, Team Leader
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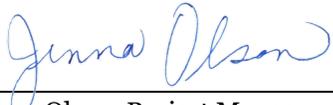


Loren Walker,
Lead CRP Quality Assurance Specialist
Quality Assurance Team

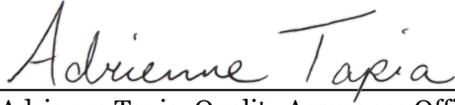
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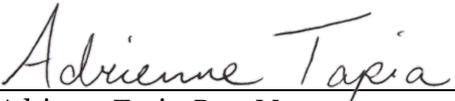
Brazos River Authority



Jenna Olson, Project Manager 8/20/25
Date



Adrienne Tapia, Quality Assurance Officer 8/20/25
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Adrienne Tapia, Data Manager 8/20/25
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Jeremy Nickolai, Field Operations Manager 20AUG25
Date

**Brazos River Authority
Environmental Services
Laboratory**



Nicole Boumans, Laboratory Manager 20Aug25
Date



Katherine Lathen,
Deputy Quality Assurance Officer 20Aug25
Date

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List of Acronyms

ALM	Aquatic Life Monitoring
ALU	Aquatic Life Use
AWRL	Ambient Water Reporting Limit
BMP	Best Management Practices
BRA	Brazos River Authority
BS	Biased Season
CAP	Corrective Action Plan
CE	Collecting Entity
CFR	Code of Federal Regulations
COC	Chain of Custody
CRP	Clean Rivers Program
DMRG	Surface Water Quality Monitoring Data Management Reference Guide
DM&A	Data Management and Analysis
DQAO	Deputy Quality Assurance Officer
EPA	United States Environmental Protection Agency
FY	Fiscal Year
GPS	Global Positioning System
HDPE	High Density Polyethylene
IBWC	International Boundary and Water Commission
IR	Integration Report
LCS	Laboratory Control Sample
LCSD	Laboratory Control Sample Duplicate
LDPE	Low Density Polyethylene
LIMS	Laboratory Information Management System
LOD	Limit of Detection
LOQ	Limit of Quantitation
MT	Monitoring Type
MS	Matrix Spike
MSD	Matrix Spike Duplicate
NELAC	National Environmental Laboratories Accreditation Conference
NELAP	National Environmental Laboratory Accreditation Program
PM	Project Manager
QA	Quality Assurance
QAM	Quality Assurance Manager
QAO	Quality Assurance Officer
QAPP	Quality Assurance Project Plan
QAS	Quality Assurance Specialist
QC	Quality Control
QM	Quality Manual
QMP	Quality Management Plan
RPD	Relative Percent Difference
RT	Routine Monitoring
SE	Submitting Entity
SLOC	Station Location
SOP	Standard Operating Procedure
SWQM	Surface Water Quality Monitoring
SWQMIS	Surface Water Quality Monitoring Information System
TAC	Texas Administrative Code
TCEQ	Texas Commission on Environmental Quality
TDS	Total Dissolved Solids
TKN	Total Kjeldahl Nitrogen
TMDL	Total Maximum Daily Load
TNI	The NELAC Institute
TP	Total Phosphorous

TSS	Total Suspended Solids
TSWQS	Texas Surface Water Quality Standards
TWDB	Texas Water Development Board
USACE	United States Army Corps of Engineers
VOA	Volatile Organic Analytes
WQS	Water Quality Standards

A4 Problem Definition/Background

In 1991, the Texas Legislature passed the Texas Clean River Act (Senate Bill 818) in response to growing concerns that water resource issues were not being pursued in an integrated, systematic manner. The act requires that ongoing water quality assessments be conducted for each river basin in Texas, an approach that integrates water quality issues within the watershed. The Clean Rivers Program (CRP) legislation mandates that each river authority (or local governing entity) shall submit quality-assured data collected in the river basin to the commission. Quality-assured data in the context of the legislation means data that comply with Texas Commission on Environmental Quality (TCEQ) rules for surface water quality monitoring (SWQM) programs, including rules governing the methods under which water samples are collected and analyzed and data from those samples are assessed and maintained. This QAPP addresses the program developed between the Brazos River Authority and the TCEQ to carry out the activities mandated by the legislation. The QAPP was developed and will be implemented in accordance with provisions of the TCEQ Quality Management Plan (QMP), Revision 30 or most recent version.

The purpose of this QAPP is to clearly delineate Brazos River Authority Quality Assurance (QA) policy, management structure, and procedures which will be used to implement the QA requirements necessary to verify and validate the surface water quality data collected. The QAPP is reviewed by the TCEQ to help ensure that data generated for the purposes described above are of known and documented quality and deemed acceptable for their intended use. This process will ensure that data collected under this QAPP and submitted to the Surface Water Quality Monitoring Information System (SWQMIS) have been collected and managed in a way that guarantees its reliability and therefore can be used in water quality assessments, total maximum daily load (TMDL) projects, water quality standards development, permit decisions, and other program activities deemed appropriate by the TCEQ. Project results will be used to support the achievement of CRP objectives, as contained in the *Guidance for Partners in the Texas Clean Rivers Program FY 2026–2027*.

In 1995, the Brazos River Authority designed the Basin Monitoring Program as the major water quality data collection effort in the Brazos River basin. The Program provides a basin-wide approach to the collection of water quality data that encourages input from local Steering Committee members. The Program is designed to provide specific types of water quality data, while providing flexibility to address dynamic water quality issues throughout the basin.

A5 Project/Task Description

This QAPP applies to routine monitoring throughout the Brazos River basin and biological/habitat assessments on selected sites.

The Clean Rivers Program for the Brazos River Basin is designed to collect water quality samples in each designated segment of the basin in concert with the TCEQ Regional personnel. The sampling is conducted on a periodic basis for water quality constituents that the TCEQ and Brazos River Authority use to assess the status of water quality. The information collected through this program is communicated to stakeholders who assist in setting priorities for monitoring locations. When the data show possible water quality concern or stakeholders indicate a concern, the CRP will focus more resources on those areas to collect water quality data and better define the water quality issue.

Sites are selected for routine monitoring and biological assessment based on stakeholder input, TCEQ assessment needs, budgetary considerations, data gaps and support of ongoing collection efforts. BRA will monitor a minimum of 93 routine stations per year. The sampling frequency and the types of parameter groups that are currently planned for collection in FY 2026 include: at a minimum, 19 monthly stations for field, select conventional parameters, and bacteria; 66 quarterly stations for field, select conventional parameters, and bacteria; five quarterly stations for field and bacteria only; seven bi-annual stations for field, select conventional and bacteria; and one station for field, 24-hr dissolved oxygen, bacteria and flow will be attempted if environmental conditions allow. The above monitoring will be routine monitoring with the objectives of collecting surface water data needed for conducting water quality assessments and identifying water quality trends. Two biological assessments will be attempted at the standard frequency of 2 if environmental conditions

allow. Biological monitoring will be performed with the objective of providing baseline data on environmental conditions and to determine if criteria for ALU and dissolved oxygen criteria are being attained.

BRA may also monitor and submit data for additional stations in-kind as there are monitoring needs in excess of available funding. If additional funding is made available during the Contract period or if the allotted budget is underutilized, the in-kind stations may then be included in the Contract and reimbursement will be allowed.

See Appendix B for the project-related work plan tasks and schedule of deliverables for a description of work defined in this QAPP.

See Appendix B for sampling design and monitoring pertaining to this QAPP.

Amendments to the QAPP

Amendments to the QAPP may be necessary to address incorrectly documented information or to reflect changes in project organization, tasks, schedules, objectives, and methods. Requests for amendments will be directed from the Brazos River Authority Project Manager (PM) to the TCEQ CRP PM electronically. The Brazos River Authority will submit a completed QAPP amendment document, including a justification of the amendment, a table of changes, and all pages, sections, and attachments affected by the amendment. Amendments are effective immediately upon approval by the Brazos River Authority PM, the Brazos River Authority Quality Assurance Officer (QAO), the TCEQ CRP PM, the TCEQ CRP Lead Quality Assurance Specialist (QAS), the TCEQ CRP Project QAS, the TCEQ CRP Team Leader, the TCEQ Data Management and Analysis (DM&A) Team Leader, and any additional parties affected by the amendment. Amendments are not retroactive. No work shall be implemented without an approved QAPP or amendment prior to the start of work. Any activities under this contract that commence prior to the approval of the governing QA document constitute a deficiency and are subject to corrective action as described in section C1 of this QAPP. Any deviation or deficiency from this QAPP which occurs after the execution of this QAPP will be addressed through a corrective action plan (CAP). An amendment may be a component of a CAP to prevent future recurrence of a deviation.

Amendments will be incorporated into the QAPP by way of attachment and distributed to personnel on the distribution list by the Brazos River Authority PM. If adherence letters are required, the Brazos River Authority will secure an adherence letter from each sub-tier project participant (e.g., subcontractors, sub-participant, or other units of government) affected by the amendment stating the organization's awareness of and commitment to requirements contained in each amendment to the QAPP. The Brazos River Authority will maintain this documentation as part of the project's QA records and ensure that the documentation is available for review.

Special Project Appendices

Projects requiring QAPP appendices will be planned in consultation with the Brazos River Authority, the TCEQ CRP PM, and TCEQ technical staff. Appendices will be written in an abbreviated format and will reference the Brazos River Authority QAPP where appropriate. Appendices will be approved by the Brazos River Authority PM, the Brazos River Authority QAO, the Laboratory (as applicable), the TCEQ CRP PM, the TCEQ CRP Project QAS, the TCEQ Lead QAS, TCEQ CRP Team Leader, the TCEQ DM&A Team Leader, and additional parties affected by the appendix, as appropriate. Copies of approved QAPP appendices will be distributed by the Brazos River Authority to project participants before data collection activities commence. The Brazos River Authority will secure written documentation from each sub-tier project participant (e.g., subcontractors, subparticipants, other units of government) stating the organization's awareness of and commitment to requirements contained in each special project appendix to the QAPP. The Brazos River Authority will maintain this documentation as part of the project's QA records and ensure that the documentation is available for review.

A6 Quality Objectives and Criteria

The purpose of routine water quality monitoring is to collect surface water quality data that can be used to characterize water quality conditions, identify significant long-term water quality trends, support water quality standards development, support the permitting process, and conduct water quality assessments in accordance with TCEQ's [Guidance for Assessing and Reporting Surface Water Quality in Texas, February 2024](https://www.tceq.texas.gov/downloads/water-quality/assessment/integrated-report-2024/2024-guidance.pdf) or most recent version (<https://www.tceq.texas.gov/downloads/water-quality/assessment/integrated-report-2024/2024-guidance.pdf>). These water quality data, and data collected by other organizations (e.g., United

States Geological Survey [USGS], TCEQ, etc.), will be subsequently reconciled for use and assessed by the TCEQ.

Systematic watershed monitoring is defined as sampling that is planned for a short duration (1 to 2 years), is designed to screen waters that would not normally be included in the routine monitoring (RT) program, investigates areas of potential concern, and investigates possible sources of water quality impairments or concerns. Due to the limitations regarding these data (e.g., not temporally representative, limited number of samples, biological sampling does not meet the specimen vouchering requirements), the data will be used to determine whether any locations have values exceeding the TCEQ's water quality criteria and/or screening levels (or in some cases values elevated above normal). The Brazos River Authority will use this information to determine future monitoring priorities. These water quality data and data collected by other organizations (e.g., USGS, TCEQ, etc.), will be subsequently reconciled for use and assessed by the TCEQ.

Brazos River Authority will conduct biological and diel monitoring using specifications found in TCEQ SOP, V1 – *Surface Water Quality Monitoring Procedures, Volume 1: Physical and Chemical Monitoring Methods*, 2012 (RG-415) and TCEQ SOP, V2 – *Surface Water Quality Monitoring Procedures, Volume 2: Methods for Collecting and Analyzing Biological Assemblage and Habitat Data*, 2014 (RG-416).

The measurement performance specifications to support the project purpose for a minimum data set are specified in Appendix A.

Ambient Water Reporting Limits (AWRLs)

For surface water to be evaluated for compliance with Texas Surface Water Quality Standards (TSWQS) and screening levels, data must be reported at or below specified reporting limits. To ensure data are collected at or below these reporting limits, required ambient water reporting limits (AWRLs) have been established. A full listing of AWRLs can be found at

<https://www.tceq.texas.gov/assets/public/waterquality/crp/QA/awrlmaster.pdf>

The limit of quantitation (LOQ) is the minimum reporting limit, concentration, or quantity of a target variable (e.g., target analyte) that can be reported with a specified degree of confidence by the laboratory analyzing the sample. Analytical results shall be reported down to the laboratory's LOQ (i.e., the laboratory's LOQ for a given parameter is its reporting limit) as specified in Appendix A.

The following requirements must be met in order to report results to the CRP:

- The laboratory's LOQ for each analyte must be set at or below the AWRL. It is the responsibility of Brazos River Authority to ensure that any laboratories used to generate CRP data have satisfactory LOQs.
- Once the LOQ is established in the QAPP, that is the reporting limit for that parameter until such time as the laboratory amends the QAPP and lists an updated LOQ.
- The laboratory must demonstrate its ability to quantitate at its LOQ for each analyte by running an LOQ check sample for each analytical batch of CRP samples analyzed.
- Under reasonable circumstances (e.g., the use of a subcontracted lab), data may be reported above or below the LOQ stated in this QAPP, so long as the LOQ remains at or below the AWRL stated in this QAPP.
- Measurement performance specifications for LOQ check samples are found in Appendix A.
- Exceptions:
 - When specific conductance reads greater than 3,000 but less than 10,000, BRA ESL uses Colilert-18 media. CRP requires dilution 1:10, requiring an LOQ of 10 MPN/100mls.

Laboratory Measurement Quality Control (QC) Requirements and Acceptability Criteria are provided in Section B4.

Precision

Precision is the degree to which a set of observations or measurements of the same property, obtained under similar conditions, conform to themselves. It is a measure of agreement among replicate measurements of the same property, under prescribed similar conditions, and is an indication of random error.

Laboratory precision is assessed by comparing replicate analyses of laboratory control samples (LCS) in the sample matrix (e.g., deionized water, sand, commercially available tissue), matrix spike/matrix spike duplicate (MS/MSD), or sample/duplicate (DUP) pairs, as applicable. Precision results are compared against measurement performance specifications and used during evaluation of analytical performance. Program-defined measurement performance specifications for precision are defined in Appendix A.

Bias

Bias is the systematic or persistent distortion of a measurement process, which causes errors in one direction (i.e., the expected sample measurement is different from the sample's true value). Bias is a statistical measurement of correctness and includes multiple components of systematic error. Bias is determined through the analysis of LCS and LOQ check samples prepared with verified and known amounts of all target analytes in the sample matrix (e.g., deionized water, sand, commercially available tissue) and by calculating percent recovery. Results are compared against measurement performance specifications and used during evaluation of analytical performance. Program-defined measurement performance specifications for bias are specified in Appendix A.

Representativeness

Site selection, the appropriate sampling regime, comparable monitoring and collection methods, and use of only approved analytical methods will assure that the measurement data represents the conditions at the site. Routine data collected under CRP are considered to be spatially and temporally representative of ambient water quality conditions. Water quality data are collected on a routine frequency and are separated by approximately even time intervals. At a minimum, samples are collected over at least two seasons (to include inter-seasonal variation) and over two years (to include inter-year variation) and include some data collected during an index period (March 15–October 15). Although data may be collected during varying regimes of weather and flow, the data sets will not be biased toward unusual conditions of flow, runoff, or season. The goal for meeting maximum representation of the water body will be tempered by funding availability.

Comparability

Confidence in the comparability of routine data sets for this project and for water quality assessments is based on the commitment of project staff to use only approved sampling and analysis methods and QA/QC protocols in accordance with quality system requirements as described in this QAPP and in TCEQ guidance. Comparability is also guaranteed by reporting data in standard units, by using accepted rules for rounding figures, and by reporting data in a standard format as specified in the Data Management Plan in Section B7.

Completeness

The completeness of the data describes how much of the data are available for use compared to the total potential data. Ideally, 100% of the data should be available. However, the possibility of unavailable data due to accidents, insufficient sample volume, broken or lost samples, etc. is to be expected. Therefore, it will be a general goal of the project(s) that 90% data completion is achieved.

A7 Distribution List

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Katherine Lathen, Deputy Quality Assurance Officer
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katherine.lathen@brazos.org

The TCEQ CRP PM will provide the approved QAPP and any amendments and appendices to TCEQ staff listed in A7 and the Brazos River Authority. The Brazos River Authority will provide copies of this project plan and any amendments or appendices of this plan to each person on this list and to each sub-tier project participant (e.g., subcontractors, subparticipants, or other units of government). The Brazos River Authority will document distribution of the plan and any amendments and appendices, maintain this documentation as part of the project's quality assurance records, and ensure the documentation is available for review.

A8 Project/Task Organization

Description of Responsibilities

TCEQ

Jason Godeaux

Manager, Monitoring and Assessment Section

Responsible for oversight of the implementation of CRP QAPPs, directs the day-to-day management of the section.

Sarah Whitley

Team Leader, Water Quality Standards and Clean Rivers Program

Responsible for TCEQ activities supporting the development and implementation of the Texas CRP. Responsible for verifying that the TCEQ QMP is followed by TCEQ CRP staff. Supervises TCEQ CRP staff. Reviews and responds to any deficiencies, corrective actions, or findings related to the area of responsibility. Oversees the development of QA guidance for the CRP. Reviews and approves all QA audits, corrective actions, reports, work plans, contracts, QAPPs, and TCEQ QMP. Enforces corrective action, as required, where QA protocols are not met. Ensures CRP personnel are fully trained.

Sunshyne Hendrix

CRP Project Quality Assurance Specialist

Serves as liaison between CRP management and TCEQ QA management. Participates in the development, approval, implementation, and maintenance of written QA standards (e.g., Program Guidance, SOPs, QAPPs, QMP). Serves on planning team for CRP special projects. Reviews and approves CRP QAPPs in coordination with other CRP staff. Coordinates documentation and monitors implementation of corrective actions for the CRP.

Sunshyne Hendrix

CRP Project Manager

Responsible for the development, implementation, and maintenance of CRP contracts. Tracks, reviews, and approves deliverables. Participates in the development, approval, implementation, and maintenance of written QA standards (e.g., Program Guidance, SOPs, QAPPs, QMP). Coordinates the review and approval of CRP QAPPs in coordination with the TCEQ CRP Project QAS. Ensures maintenance of QAPPs. Assists TCEQ CRP Lead QAS in conducting Brazos River Authority audits. Verifies QAPPs are being followed by contractors and that projects are producing data of known quality. Coordinates project planning with the Brazos River Authority PM. Reviews and approves data and reports produced by contractors. Notifies TCEQ CRP QA Specialists of circumstances that may adversely affect the quality of data derived from the collection and analysis of samples. Develops, enforces, and monitors corrective action measures to ensure contractors meet deadlines and scheduled commitments.

Cathy Anderson

Team Leader, Data Management and Analysis Team

Participates in the development, approval, implementation, and maintenance of written QA standards (e.g., Program Guidance, SOPs, QAPPs, QMP). Ensures DM&A staff perform data management-related tasks.

Scott Delgado

CRP Data Manager, Data Management and Analysis Team

Responsible for coordination and tracking of CRP data sets from initial submittal through TCEQ CRP PM review and approval. Ensures that data are reported following instructions in the Data Management Reference Guide (DMRG), July 2019 or most current version. Runs automated data validation checks in SWQMIS and coordinates data verification and error correction with TCEQ CRP PMs. Generates SWQMIS summary reports to assist CRP PMs' data review. Identifies data anomalies and inconsistencies. Provides training and guidance to CRP and planning agencies on technical data issues to ensure that data are submitted according to documented

procedures. Reviews QAPPs for valid stream monitoring stations. Checks validity of parameter codes, submitting entity (SE) code(s), collecting entity (CE) code(s), and monitoring type (MT) code(s). Develops and maintains data management-related SOPs for CRP data management. Coordinates and processes data correction requests. Participates in the development, implementation, and maintenance of written QA standards (e.g., Program Guidance, SOPs, QAPPs, QMP).

D. Jody Koehler

TCEQ Quality Assurance Manager

Responsible for coordinating development and implementation of TCEQ's QA program. Provides oversight and guidance for TCEQ's QA program. Responsible for the development and maintenance of the TCEQ QMP. TCEQ's QA Manager, or designated QA staff in the Laboratory and Quality Assurance Section of the Air Monitoring Division, is responsible for review and approval of program/project QAPPs to ensure QAPPs conform to applicable requirements as detailed in TCEQ's QMP.

Loren Walker

CRP Lead Quality Assurance Specialist

Participates in the development, approval, implementation, and maintenance of written QA standards (e.g., Program Guidance, SOPs, QAPPs, QMP). Assists program manager and TCEQ CRP Project QAS in developing and implementing the quality system. Reviews and approves CRP QAPPs, QAPP amendments, and QAPP special appendices. Prepares and distributes annual audit plans. Conducts monitoring systems audits of planning agencies. Concurs with corrective actions. Conveys QA problems to appropriate management. Recommends that work be stopped in order to safeguard programmatic objectives, worker safety, public health, or environmental protection. Ensures maintenance of audit records for the CRP.

BRAZOS RIVER AUTHORITY

Jenna Olson

Brazos River Authority Project Manager

Responsible for implementing and monitoring CRP requirements in contracts, QAPPs, and QAPP amendments and appendices. Coordinates basin planning activities and work of basin partners. Ensures monitoring systems audits are conducted to ensure QAPPs are followed by Brazos River Authority participants and that projects are producing data of known quality. Ensures that subparticipants are qualified to perform contracted work. Ensures TCEQ CRP PM and/or QA Specialists are notified of deficiencies and corrective actions, and that issues are resolved. Responsible for validating that data collected are acceptable for reporting to the TCEQ.

Adrienne Tapia

Brazos River Authority Quality Assurance Officer

Responsible for coordinating the implementation of the QA program. Responsible for writing and maintaining the QAPP and monitoring its implementation. Responsible for maintaining records of QAPP distribution, including appendices and amendments. Responsible for maintaining written records of sub-tier commitment to requirements specified in this QAPP. Responsible for identifying, receiving, and maintaining project QA records. Responsible for coordinating with the TCEQ CRP PM to resolve QA-related issues. Notifies the Brazos River Authority PM of particular circumstances that may adversely affect the quality of data. Coordinates and monitors deficiencies and corrective action. Coordinates and maintains records of data verification and validation. Coordinates the research and review of technical QA material and data related to water quality monitoring system design and analytical techniques. Conducts monitoring systems audits on project participants to determine compliance with project and program specifications, issues written reports, and follows through on findings.

Adrienne Tapia

Brazos River Authority Data Manager

Responsible for ensuring that field data are properly reviewed and verified. Responsible for the transfer of basin quality-assured water quality data to the TCEQ in a format compatible with SWQMIS. Maintains quality-assured data on Brazos River Authority internet sites.

Jeremy Nickolai

Brazos River Authority Field Operations Manager

Responsible for coordinating field activities to ensure correctness, completeness, compliance, and consistency with project goals. Responsible for supervision of Aquatic Scientists. Ensures that field staff is properly trained and that training records are maintained.

Nicole Boumans

Brazos River Authority Laboratory Manager

Responsible for initial review and verification of lab data for correctness, completeness, compliance, and consistency with project goals. Coordinates daily lab function. Supervises Laboratory Analysts.

Katherine Lathen

Brazos River Authority Deputy Quality Assurance Officer

Assists with the coordination and implementation of the QA program. Responsible for ensuring that field and lab data from the BRA Environmental Services Laboratory are properly reviewed and verified for compliance with BRA SOPs, CRP QAPP, and NELAP. Coordinates and maintains records of data verification and validation. Maintains the daily corrective action process. Conducts laboratory internal audits as detailed in BRA SOPs. Assists with writing and maintaining the QAPP. Responsible for the transfer of basin quality-assured water quality data to the TCEQ in a format compatible with SWQMIS.

A9 Project QAM Independence

TCEQ uses a semi-decentralized QA program, which is organizationally independent of operational programs and activities within the agency. TCEQ's QA program has sufficient access and authority to coordinate the development and implementation of the agency's quality system.

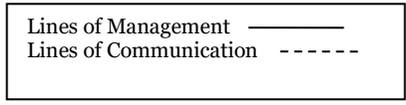
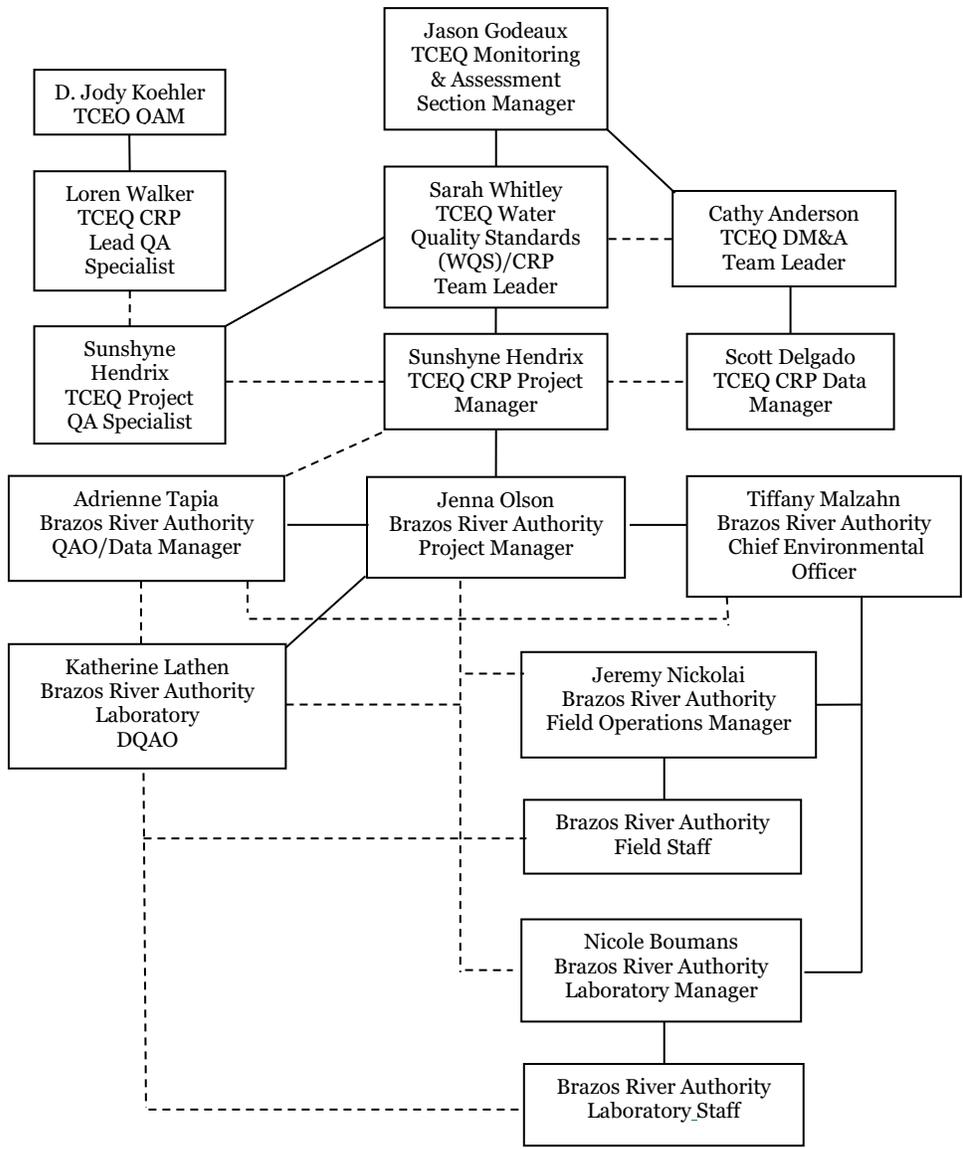
The TCEQ QA Manager (QAM) and designated TCEQ QA staff from the Laboratory and Quality Assurance Section within the Air Monitoring Division of the Office of Air are independent of activities performed by CRP. No CRP staff have authority to sign QAPPs, amendments, or appendices on behalf of TCEQ's QAM or the Lead CRP QAS. Similarly, TCEQ's QAM and the Lead CRP QAS cannot sign QAPPs, amendments or appendices on behalf of CRP staff.

Roles of project QA staff are described in Section A8. An illustration of QA independence and lines of communication and supervision for this project are detailed in the project organization chart in A10. Communication for deficiencies and corrective actions are described in Section C1.

A10 Project Organizational Chart and Communication

Project Organization Chart

Figure A10.1. Organization Chart with Lines of Communication



A11 Special Training/Certification

Before new field personnel independently conduct field work, a Senior Aquatic Scientist or the Field Operations Manager trains them in proper instrument calibration, field sampling techniques, and field analysis procedures. The Brazos River Authority QAO (or designee) will document the successful field demonstration. The Brazos River Authority QAO (or designee) will retain documentation of training and the successful field demonstration in the employee's personnel file (or other designated location) and ensure that the documentation will be available during monitoring systems audits.

The requirements for obtaining certified positional data using a global positioning system (GPS) are located in Section B7, Data Management.

Contractors and subcontractors must ensure that laboratories analyzing samples under this QAPP meet the requirements contained in The National Environmental Laboratories Accreditation Conference (NELAC) Institute Standard (2016) Volume 1, Module 2, Section 4.5 (concerning Subcontracting of Environmental Tests).

A12 Documents and Records

The documents and records that describe, specify, report, or certify activities are listed. The list below is limited to documents and records that may be requested for review during a monitoring systems audit.

Table A12.1 Project Documents and Records

Document/Record	Location	Retention (yrs)	Format
QAPPs, amendments and appendices	Brazos River Authority	10+number of years since most recent IR*	Electronic
Field SOPs	Brazos River Authority	10+number of years since most recent IR*	Electronic
Laboratory quality manuals	Brazos River Authority/ Laboratory(ies)	10+number of years since most recent IR*	Electronic
Laboratory SOPs	Brazos River Authority/ Laboratory(ies)	10+number of years since most recent IR*	Electronic
QAPP distribution documentation	Brazos River Authority	5 past end of contract period	Electronic
Field staff training records	Brazos River Authority	5 past end of contract period	Electronic
Field equipment calibration/maintenance logs	Brazos River Authority	5 past end of contract period	Electronic
Field instrument printouts	Brazos River Authority	5 past end of contract period	Electronic
Field notebooks or data sheets	Brazos River Authority	5 past end of contract period	Paper/Electronic
Chain of custody records	Brazos River Authority	5 past end of contract period	Paper/Electronic
Laboratory calibration records	Laboratory(ies)	5 past end of	Electronic

		contract period	
Laboratory instrument printouts	Laboratory(ies)	5 past end of contract period	Electronic
Laboratory data reports/results	Brazos River Authority/ Laboratory(ies)	5 past end of contract period	Electronic
Laboratory equipment maintenance logs	Laboratory(ies)	5 past end of contract period	Electronic
Corrective action documentation	Brazos River Authority/ Laboratory(ies)	5 past end of contract period	Electronic

*For example, the most recent IR is from 2024; thus, records must be kept for 11 years.

Laboratory Test Reports

Test/data reports from the laboratory must document the test results clearly and accurately. Routine data reports should be consistent with The NELAC Institute (TNI) Standard (2016), Volume 1, Module 2, Section 5.10 and include the information necessary for the interpretation and validation of data. The requirements for reporting data and the procedures are provided.

In order to achieve a paperless laboratory, all data are entered directly into the laboratory information management system (LIMS) including the following:

- Sample results including analyst, date/time of analysis, and method used.
- Units of measurement
- Sample matrix
- Dry weight or wet weight (as applicable)
- Station information
- Date and time of collection
- Sample depth
- Holding time for *E. coli*
- LOQ and limit of detection (LOD)
- Certification of NELAP compliance

All data required necessary for review, verification, and interpretation of data are contained in the BRA laboratory database.

Data reports are only generated upon request. At the very minimum, test reports include the information listed above.

Electronic Data

Data will be submitted electronically to the TCEQ in the event/result file format described in the most current version of the [DMRG](https://www.tceq.texas.gov/waterquality/data-management/dmrg_index.html), which can be found at https://www.tceq.texas.gov/waterquality/data-management/dmrg_index.html. A completed data review checklist and data summary (see Appendix F) will be included with each data submittal.

Biological data will be submitted electronically to the TCEQ in the ASCII Pipe-Delimited Event text file, ASCII Pipe-Delimited Result text file, README.txt file, and BLOB file format described Chapter 12 TCEQ Partner Agencies and Contractors paragraphs in the most current version of the [DMRG](https://www.tceq.texas.gov/waterquality/data-management/dmrg_index.html), which can be found at https://www.tceq.texas.gov/waterquality/data-management/dmrg_index.html.

B1 Sampling Process Design

See Appendix B for sampling process design information and monitoring tables associated with data collected under this QAPP.

B2 Sampling Methods

Field Sampling Procedures

Field sampling will be conducted in accordance with the latest versions of the *TCEQ Surface Water Quality Monitoring Procedures Volume 1: Physical and Chemical Monitoring Methods, 2012* (RG-415) and *Volume 2: Methods for Collecting and Analyzing Biological Assemblage and Habitat Data, 2014* (RG-416), collectively referred to as “SWQM Procedures.” Updates to SWQM Procedures are posted to the Surface Water Quality Monitoring Procedures website (https://www.tceq.texas.gov/waterquality/monitoring/swqm_guides.html), and shall be incorporated into the Brazos River Authority’s procedures, QAPP, SOPs, etc., within 60 days of any final published update. Additional aspects outlined in Section B below reflect specific requirements for sampling under CRP and/or provide additional clarification.

Table B2.1 Sample Storage, Preservation, and Handling Requirements

Parameter	Matrix	Container	Sample Volume	Preservation	Holding Time	
Chloride	Water	Nalgene Bottle	125 ml ¹	Cool to >0 and <6°C	28 days	
Sulfate					48 hours	
Nitrate Nitrogen						
Nitrite Nitrogen						
Orthophosphate Phosphorous						48 hours Filter within 15 minutes of collection ² Use Parameter code 00671
Orthophosphate Phosphorous Lab Filtered						48 hours If not filtered within 15 minutes of collection, use Parameter code 70507
<i>E. coli</i>	Plastic (sterile)	100 ml or 250 ml (duplicates)	Sodium Thiosulfate Cool to >0 and <6°C	8 hours ³		
Enterococcus				8 hours		
Total suspended solids (TSS)	LDPE Cubitainer	1 L	Cool to >0 and <6°C	168 hours		
Total dissolved solids (TDS)	Brown Nalgene Bottle	500 ml ⁴		48 hours		
Turbidity						
Chlorophyll a					Dark, ice, cool to >0 and <6°C before filtration Dark, frozen after filtration	≤48 hours unfiltered 24 days filtered
Total Phosphorous (TP)	Nalgene Bottle	250 ml ⁵	Cool to >0 and <6°C	28 days		

Total Kjeldahl Nitrogen (TKN)			H ₂ SO ₄ to pH<2	
Ammonia				
Benthic Macro Invertebrates	Glass	Variable ⁶	10% formalin in field 70% ethanol after washing	Permanent preservation in ethanol
Fish			10% formalin in field 70% ethanol	7 days in formalin Permanent preservation in ethanol

¹Ion chromatograph analytes (chloride, nitrite nitrogen, nitrate nitrogen, orthophosphate-phosphorus and sulfate) will be taken from the same 125 mL sample after filtration of orthophosphate phosphorus.

²Preservation of orthophosphate phosphorus is performed immediately upon collection or within 15 minutes of collection by filtration in the field.

³*E. coli* samples should always be processed as soon as possible and incubated no later than 8 hours from time of collection. When transport conditions necessitate sample incubation after 8 hours from time of collection, the holding time may be extended, and samples must be processed as soon as possible and within 30 hours.

⁴TDS, Turbidity, and Chlorophyll *a* are analyzed from a single 500 mL sample for a given monitoring location.

⁵TKN, TP, and Ammonia are analyzed from a single 250 mL sample for each monitoring location.

⁶Sample volume is dependent on number of organisms collected.

Sample Containers

Certificates from sample container manufacturers are maintained in LIMS and on Environmental Services SharePoint by the Brazos River Authority. Sample containers used for conventional parameters are purchased pre-cleaned and are disposable from Quality Environmental Containers.

- IDEXX sterile 120 and 290 mL bottles are used for bacteria sampling.
- 1L LDPE Cubitainers are used for TSS sampling.
- 500 mL Brown HDPE bottles are used for chlorophyll *a*, TDS, and Turbidity sampling.
- 250 mL White HDPE bottles are used for TKN, TP, and ammonia sampling.
- 125 mL White HDPE bottles are used for chloride, sulfate, nitrate nitrogen, nitrite nitrogen, and orthophosphate phosphorous sampling.

Processes to Prevent Contamination

SWQM Procedures outline the necessary steps to prevent contamination of samples, including: direct collection into sample containers, when possible; use of certified containers for organics; and clean sampling techniques for metals. Field QC samples (identified in Section B4) are collected to verify that contamination has not occurred.

Documentation of Field Sampling Activities

Field sampling activities are documented on field data sheets as presented in Appendix D. Flow worksheets, aquatic life use monitoring checklists, habitat assessment forms, field biological assessment forms, and records of bacteriological analyses (if applicable) are part of the field data record. The following will be recorded for all visits:

- Station ID
- Sampling date
- Location
- Sampling depth
- Sampling time
- Sample collector's initials
- Values for all field parameters collected

Additional notes containing detailed observational data not captured by field parameters may include:

- Water appearance
- Weather
- Biological activity
- Recreational activity
- Unusual odors
- Pertinent observations related to water quality or stream uses
- Watershed or instream activities
- Specific sample information
- Missing parameters

Recording Data

For the purposes of this section and subsequent sections, all field and laboratory personnel follow the basic rules for recording information as documented below:

- Write legibly, in indelible ink.
- Make changes by crossing out original entries with a single line strike-out, entering the changes, and initialing and dating the corrections.
- Close-out incomplete pages with an initialed and dated diagonal line.

Sampling Method Requirements or Sampling Process Design Deficiencies, and Corrective Action

Examples of sampling method requirements or sample design deficiencies include but are not limited to such things as inadequate sample volume due to spillage or container leaks, failure to preserve samples appropriately, contamination of a sample bottle during collection, storage temperature and holding time exceedance, sampling at the wrong site, etc. Any deviations from the QAPP, SWQM Procedures, or appropriate sampling procedures may invalidate data and require documented corrective action. Corrective action may include for samples to be discarded and re-collected. It is the responsibility of the Brazos River Authority PM, in consultation with the Brazos River Authority QAO, to ensure that the actions and resolutions to the problems are documented and that records are maintained in accordance with this QAPP. In addition, these actions and resolutions will be conveyed to the TCEQ CRP PM both verbally and in writing in the project progress reports and by completion of a CAP.

The definition of and process for handling deficiencies and corrective action are defined in Section C1.

Analytical Methods

The analytical methods, associated matrices, and performing laboratories are listed in Appendix A. The authority for analysis methodologies under CRP is derived from the Texas Administrative Code (TAC), Title 30, Chapter 307, in that data generally are generated for comparison to those standards and/or criteria. The TSWQS state “procedures for laboratory analysis must be in accordance with the most recently published edition of the book entitled Standard Methods for the Examination of Water and Wastewater, the TCEQ SWQM Procedures as amended, 40 Code of Federal Regulations (CFR) 136, or other reliable procedures acceptable to the TCEQ, and in accordance with chapter 25 of this title.”

Laboratories collecting data under this QAPP must be accredited by the National Environmental Laboratory Accreditation Program (NELAP) in accordance with TAC, Title 30, Chapter 25. Copies of laboratory quality manuals (QMs) and SOPs shall be made available for review by the TCEQ.

Standards Traceability

All standards used in the field and laboratory are traceable to certified reference materials. Standards

preparation is fully documented and maintained in the BRA LIMS. Each documentation includes information concerning the standard identification, starting materials, including concentration, amount used and lot number; date prepared, expiration date and preparer's initials/signature. The reagent bottle is labeled in a way that will trace the reagent back to preparation.

Analytical Method Deficiencies and Corrective Actions

Deficiencies in field and laboratory measurement systems involve, but are not limited to such things as instrument malfunctions, failures in calibration, blank contamination, quality control samples outside QAPP-defined limits, etc. In many cases, the field technician or lab analyst will be able to correct the problem. If the problem is resolvable by the field technician or lab analyst, then they will document the problem on the field data sheet or laboratory record and complete the analysis. If the problem is not resolvable, then it is conveyed to the applicable supervisor, who will make the determination and notify the Brazos River Authority QAO if the problem compromises sample results. If the analytical system failure may compromise the sample results, the resulting data will not be reported to the TCEQ. The nature and disposition of the problem is reported on the data report which is sent to the Brazos River Authority PM. If a CAP is necessary (Figure C1.1), the Brazos River Authority QAO will submit the CAP to the TCEQ CRP PM in a timely manner for review. Additionally, the Brazos River Authority PM will summarize the CAP in the associated progress report submitted to the TCEQ CRP PM.

The definition of and process for handling deficiencies and corrective action are explained in detail in Section C1.

The TCEQ has determined that analyses associated with qualifier codes (e.g., "holding time exceedance," "sample received unpreserved," "estimated value") may have unacceptable measurement uncertainty associated with them. This will immediately disqualify analyses from submittal to SWQMIS. Therefore, data with these types of problems should not be reported to the TCEQ. Additionally, any data collected or analyzed by means other than those stated in the QAPP, or data suspect for any reason should not be submitted for loading and storage in SWQMIS. However, when data is lost, its absence will be described in the data summary report submitted with the corresponding data set, and a CAP (as described in Section C1) may be necessary.

Acquired Data

Non-directly measured data, secondary data, or acquired data involves the use of data collected under another project and collected with a different intended use than this project. The acquired data still meets the quality requirements of this project and is defined below. The following data source(s) will be used for this project:

USGS gage station data will be used throughout this project to aid in determining gage height and flow. Rigorous QA checks are completed on gage data by the USGS and the data are approved by the USGS and permanently stored at the USGS. This data will be submitted to the TCEQ under parameter code 00061 (instantaneous flow).

Reservoir stage data are collected every day from the USGS, International Boundary and Water Commission (IBWC), and the United States Army Corps of Engineers (USACE) websites. These data are preliminary and subject to revision. The Texas Water Development Board (TWDB) derives reservoir storage (in acre-feet) from these stage data (elevation in feet above mean sea level), by using the latest rating curve datasets available. These data are published at the TWDB website at <http://waterdatafortexas.org/reservoirs/statewide>. Information about measurement methodology can be found on the TWDB website. These data will be submitted to the TCEQ under parameter code 00052 (reservoir stage) and parameter code 00053 (reservoir percent full).

B3 Sample Handling and Custody

Sample Tracking

Proper sample handling and custody procedures ensure the custody and integrity of samples beginning at the

time of sampling and continuing through transport, sample receipt, preparation, and analysis.

A sample is in custody if it is in actual physical possession or in a secured area that is restricted to authorized personnel. The chain of custody (COC) form is a record that documents the possession of the samples from the time of collection to receipt in the laboratory. The following information concerning the sample is recorded on the COC form. BRA uses the field data sheet as the COC for routine sampling (See Appendix D). The following list of items matches the COC form in Appendix D.

- Date and time of collection
- Site identification
- Sample matrix
- Number of containers
- Preservative used
- Was the sample filtered
- Analyses required
- Name of collector
- Custody transfer signatures and dates and time of transfer
- Bill of lading, if applicable

Sample Labeling

Samples from the field are labeled on the container, or on a label, with an indelible marker. Label information includes:

- LIMS sample number (includes the station ID and matrix)
- Date and time of collection
- Sample depth
- Initials of collector

Sample Handling

Samples are collected in the field, filtered or acid preserved as necessary, and stored in coolers on ice. Samples are delivered to the Authority's water quality laboratory in coolers with field data sheets (COC Forms) attached. The laboratory staff examines each sample container for anomalies and ensures that all container information matches the information on the appropriate field data sheet. If the information is present and correct, the lab staff will receive the samples by signing the field data sheet "received by" block and entering the samples into the LIMS. At this instant, the samples become the responsibility of the Authority's water quality laboratory. In some instances, bacteria samples may arrive at the lab after hours. In this case, field staff will assess and record the condition of the samples on the field data sheets (COC) before analysis.

Internal sample handling, custody, and storage procedures for laboratory are described in the Brazos River Authority's Environmental Laboratory Quality Manual and Sample Receipt SOP BRA QUAL 009.

Sample Tracking Procedure Deficiencies and Corrective Action

All deficiencies associated with COC procedures, as described in this QAPP, are immediately reported to the Brazos River Authority Deputy Quality Assurance Officer (DQAO) and PM. These include such items as delays in transfer resulting in holding time violations; violations of sample preservation requirements; incomplete documentation, including signatures; possible tampering of samples; broken or spilled samples; etc. The Brazos River Authority PM, in consultation with the Brazos River Authority QAO and DQAO, will determine if the procedural violation may have compromised the validity of the resulting data. Any failures that have reasonable potential to compromise data validity will invalidate data and the sampling event should be repeated. The resolution of the situation will be reported to the TCEQ CRP PM in the project progress report. CAPs will be prepared by the Brazos River Authority and submitted to TCEQ CRP PM.

The definition of and process for handling deficiencies and corrective action are defined in Section C1.

B4 Quality Control

Sampling Quality Control Requirements and Acceptability Criteria

The minimum field QC requirements, and program-specific laboratory QC requirements, are outlined in SWQM Procedures. No field QC samples are collected since BRA does not sample for metals or organics under this QAPP.

Laboratory Measurement Quality Control Requirements and Acceptability Criteria

Batch

A batch is defined as environmental samples that are prepared and/or analyzed together with the same process and personnel, using the same lot(s) of reagents. A preparation batch is composed of one to 20 environmental samples of the same NELAP-defined matrix, meeting the above-mentioned criteria and with a maximum time between the start of processing of the first and last sample in the batch to be 24 hours. An analytical batch is composed of prepared environmental samples (extract, digestates, or concentrates) which are analyzed together as a group. An analytical batch can include prepared samples originating from various environmental matrices and can exceed 20 samples.

Method Specific QC requirements

QC samples, other than those specified later in this section (e.g., sample duplicates, surrogates, internal standards, continuing calibration samples, interference check samples, positive control, negative control, and media blank), are run as specified in the methods and in SWQM Procedures. The requirements for these samples, their acceptance criteria or instructions for establishing criteria, and corrective actions are method-specific.

Detailed laboratory QC requirements and corrective action procedures are contained within the laboratory QM and SOPs. The minimum requirements that all participants abide by are stated below.

Comparison Counting

For routine bacteriological samples, repeat counts on one or more positive samples are required, at least monthly. If possible, the analyst will compare counts with another analyst who also performs the analysis. Replicate counts by the same analyst should agree within 5 percent, and those between analysts should agree within 10 percent. The analyst(s) will record the results.

Limit of Quantitation (LOQ)

The laboratory will analyze a calibration standard (if applicable) at the LOQ published in Appendix A of this QAPP on each day calibrations are performed. In addition, an LOQ check sample will be analyzed with each analytical batch. Calibrations including the standard at the LOQ listed in Appendix A will meet the calibration requirements of the analytical method, or corrective action will be implemented.

LOQ Check Sample

An LOQ check sample consists of a sample matrix (e.g., deionized water, sand, commercially available tissue) free from the analytes of interest spiked with verified known amounts of analytes or a material containing known and verified amounts of analytes. It is used to establish intra-laboratory bias to assess the performance of the measurement system at the lower limits of analysis. The LOQ check sample is spiked into the sample matrix at a level less than or equal to the LOQ published in Appendix A of this QAPP, for each analyte for each analytical batch of CRP samples run. If it is determined that samples have exceeded the high range of the calibration curve, samples should be diluted or run on another curve. For diluted or high concentration samples run on batches with calibration curves that do not include the LOQ published in Appendix A of this QAPP, a check sample will be run at the low end of the calibration curve.

The LOQ check sample is carried through the complete preparation and analytical process and is performed at a rate of one per analytical batch.

The percent recovery of the LOQ check sample is calculated using the following equation in which %R is percent recovery, S_R is the sample result, and S_A is the reference concentration for the check sample:

$$\%R = S_R / S_A \times 100$$

Measurement performance specifications are used to determine the acceptability of LOQ check sample analyses as specified in Appendix A of this QAPP.

Laboratory Control Sample (LCS)

An LCS consists of a sample matrix (e.g., deionized water, sand, commercially available tissue) free from the analytes of interest spiked with verified known amounts of analytes or a material containing known and verified amounts of analytes. It is used to establish intra-laboratory bias to assess the performance of the measurement system. The LCS is spiked into the sample matrix at a level less than or near the midpoint of the calibration for each analyte. In cases of test methods with very long lists of analytes, LCSs are prepared with all the target analytes and not just a representative number, except in cases of organic analytes with multipeak responses.

The LCS is carried through the complete preparation and analytical process and is performed at a rate of one per preparation batch.

Results of LCSs are calculated by percent recovery (%R), which is defined as 100 times the measured concentration, divided by the true concentration of the spiked sample.

The following formula is used to calculate percent recovery, where %R is percent recovery; S_R is the measured result; and S_A is the true result:

$$\%R = S_R / S_A \times 100$$

Measurement performance specifications are used to determine the acceptability of LCS analyses as specified in Appendix A.

Laboratory Duplicates

A laboratory duplicate is an aliquot taken from the same container as an original sample under laboratory conditions and processed and analyzed independently. A laboratory duplicate is achieved by preparing 2 separate aliquots of a sample, LCS, or matrix spike. Both samples are carried through the entire preparation and analytical process. Laboratory duplicates are used to assess precision and are performed at a rate of one per preparation batch.

For most parameters except bacteria, precision is evaluated using the relative percent difference (RPD) between duplicate results as defined by 100 times the difference (range) of each duplicate set, divided by the average value (mean) of the set. For duplicate results, X_1 and X_2 , the RPD is calculated from the following equation:

$$RPD = \frac{|X_1 - X_2|}{\left(\frac{X_1 + X_2}{2}\right)} \times 100$$

If the precision criterion is exceeded, the data are not acceptable for use under this project and are not reported to TCEQ. Results from all samples associated with that failed duplicate (usually a maximum of 10 samples) are considered to have excessive analytical variability and are qualified as not meeting project QC requirements.

For bacteriological parameters, precision is evaluated using the results from laboratory duplicates. Bacteriological duplicates are analyzed at a 10% frequency (or once per preparation batch, whichever is more frequent). Sufficient volume should be collected to analyze laboratory duplicates from the same sample container.

The base-10 logarithms of the results from the original sample and its duplicate are calculated. The absolute

value of the difference between the two base-10 logarithms is calculated and compared to the precision criterion in Appendix A.

$$|\text{Log A} - \text{Log B}| = \text{Log Range}$$

If the difference in logarithms is greater than the precision criterion, the data are not acceptable for use under this project and are not reported to TCEQ. Results from all samples associated with that failed duplicate (usually a maximum of 10 samples) are considered to have excessive analytical variability and are qualified as not meeting project QC requirements.

The precision criterion in Appendix A for bacteriological duplicates applies only to samples with concentrations >10 MPN.

Matrix spike

Matrix spikes are prepared by adding a known quantity of target analyte to a specified amount of matrix sample for which an independent estimate of target analyte concentration is available.

Matrix spikes indicate the effect of the sample on the precision and accuracy of the results generated using the selected method. Matrix-specific QC samples indicate the effect of the sample matrix on the precision and accuracy of the results generated using the selected method. The information from these controls is sample/matrix specific and would not normally be used to determine the validity of the entire batch. The frequency of matrix spikes is specified by the analytical method, or a minimum of one per preparation batch, whichever is greater. To the extent possible, matrix spikes prepared and analyzed over the course of the project should be performed on samples from different sites.

The components to be spiked shall be as specified by the mandated analytical method. The results from matrix spikes are primarily designed to assess the validity of analytical results in a given matrix and are expressed as percent recovery (%R).

The percent recovery of the matrix spike is calculated using the following equation, where %R is percent recovery, S_{SR} is the concentration measured in the matrix spike, S_R is the concentration in the parent sample, and S_A is the concentration of analyte that was added:

$$\%R = \frac{S_{SR} - S_R}{S_A} \times 100$$

Matrix spike recoveries are compared to the same acceptance criteria established for the associated LCS recoveries, rather than the matrix spike recoveries published in the mandated test method. The EPA 1993 methods (i.e., ammonia-nitrogen, ion chromatography, TKN) that establish matrix spike recovery acceptance criteria are based on recoveries from drinking water that has very low interferences and variability and do not represent the matrices sampled in the CRP. If the matrix spike results are outside laboratory-established criteria, there will be a review of all other associated quality control data in that batch. If all of the quality control data in the associated batch passes, it will be the decision of the laboratory QAO or Brazos River Authority PM to report the data for the analyte that failed in the parent sample to TCEQ or to determine that the result from the parent sample associated with that failed matrix spike is considered to have excessive analytical variability and does not meet project QC requirements. Depending on the similarities in composition of the samples in the batch, the Brazos River Authority may consider excluding all of the results in the batch related to the analyte that failed recovery.

Method blank

A method blank is a sample of matrix similar to the batch of associated samples (when available) that is free from the analytes of interest and is processed simultaneously with and under the same conditions as the samples through all steps of the analytical procedures, and in which no target analytes or interferences are present at concentrations that impact the analytical results for sample analyses. The method blank is used to document contamination from the analytical process. The analysis of method blanks should yield values less than the LOQ. For very high-level analyses, the blank value should be less than 5% of the lowest value of the batch, or corrective action will be implemented. Samples associated with a contaminated blank shall be evaluated as to the best

corrective action for the samples (e.g., reprocessing, data qualifying codes). In all cases, the corrective action must be documented.

The method blank shall be analyzed at a minimum of one per preparation batch. In those instances for which no separate preparation method is used (e.g., VOA) the batch shall be defined as environmental samples that are analyzed together with the same method and personnel, using the same lots of reagents, not to exceed the analysis of 20 environmental samples.

Quality Control or Acceptability Requirements, Deficiencies, and Corrective Actions

Sampling QC excursions are evaluated by the Brazos River Authority PM, in consultation with the Brazos River Authority QAO. In that differences in sample results are used to assess the entire sampling process, including environmental variability, the arbitrary rejection of results based on pre-determined limits is not practical. Therefore, the professional judgment of the Brazos River Authority PM and QAO will be relied upon in evaluating results.

Notations of blank contamination are noted in the data summaries that accompany data deliverables.

Laboratory measurement quality control failures are evaluated by the laboratory staff. The disposition of such failures and the nature and disposition of the failure is reported to the Laboratory DQAO. The Laboratory DQAO will discuss the failure with the Brazos River Authority PM. If applicable, the Brazos River Authority PM will include this information in a CAP and submit the CAP to the TCEQ CRP PM.

The definition of and process for handling deficiencies and corrective action are defined in Section C1.

Additionally, in accordance with CRP requirements and the TNI Standard (Volume 1, Module 2, Section 4.5, Subcontracting of Environmental Tests) when a laboratory that is a signatory of this QAPP finds it necessary and/or advantageous to subcontract analyses, the laboratory that is the signatory on this QAPP must ensure that the subcontracting laboratory is NELAP-accredited (when required) and understands and follows the QA/QC requirements included in this QAPP. This includes confirming that the sub-contracting laboratory has LOQs at or below TCEQ AWRLs and performs all required QC analysis outlined in this QAPP. The signatory laboratory is also responsible for QA of the data prior to delivering it to the Brazos River Authority, including review of all applicable QC samples related to CRP data. As stated in section 4.5.5 of the TNI Standard, the laboratory performing the subcontracted work shall be indicated in the final report and the signatory laboratory shall make a copy of the subcontractor's report available to the client (Brazos River Authority) when requested.

B5 Instrument/Equipment Calibration, Testing, Inspection, and Maintenance

All sampling equipment testing and maintenance requirements are detailed in the SWQM Procedures. Sampling equipment is inspected and tested upon receipt and is assured appropriate for use by field staff. Equipment records are kept on all field equipment and a supply of critical spare parts is maintained.

All laboratory tools, gauges, instrument, and equipment testing and maintenance requirements are contained within laboratory QM.

Instrument Calibration and Frequency

Field equipment calibration requirements are contained in the SWQM Procedures. Post-calibration check error limits and the disposition resulting from errors are adhered to. Data collected from field instruments that do not meet the post-calibration check error limits specified in the SWQM Procedures will not be submitted for inclusion into SWQMIS.

Detailed laboratory calibrations are contained within the QM or SOPs.

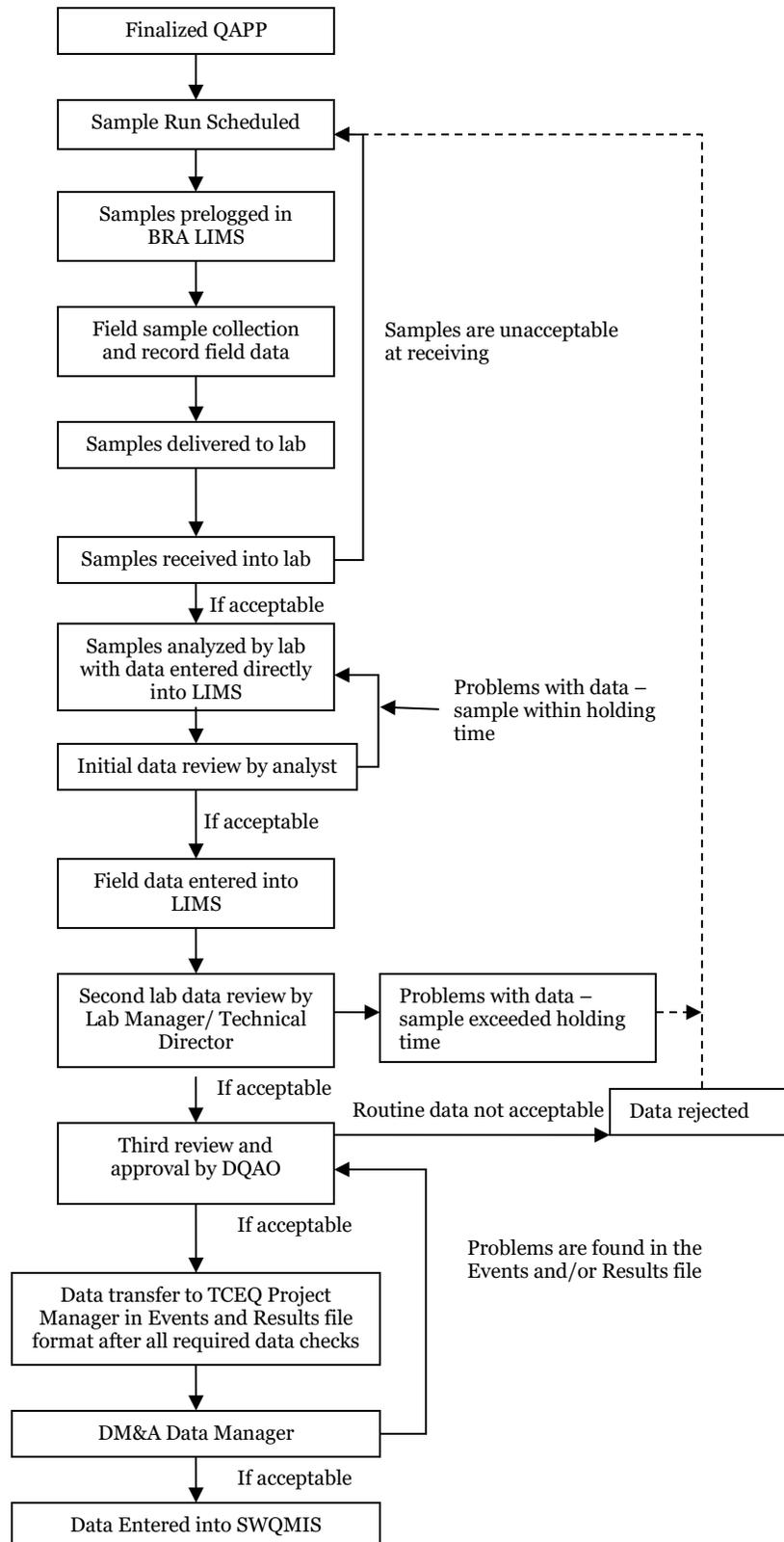
B6 Inspection/Acceptance of Supplies and Consumables

No special requirements for acceptance are specified for field sampling supplies and consumables. Procedures for acceptance of laboratory supplies and consumables are contained within the QM.

B7 Data Management

Data Management Process

Figure B7.1



Data Dictionary

Terminology and field descriptions are included in the 2019 DMRG, or most recent version.

A table outlining the entities that will be used when submitting data under this QAPP is included below to verify the entity codes included in this QAPP.

Name of Entity	Tag Prefix	Submitting Entity	Collecting Entity
Brazos River Authority	BR	BR	BR

Data Errors and Loss

Laboratory analysts review all data after data entry, constituting the first data review. If needed and the sample is still within holding time, the technician will reanalyze samples not meeting QA requirements. The Laboratory Manager or designee reviews all laboratory data following analysis and checks for calculation errors or data entry errors, constituting the second data review. The DQAO performs a third review of all data to determine validity within this QAPP. The BRA LIMS records all data corrections in an audit trail.

Record Keeping and Data Storage

All electronic records are backed-up weekly. Access to protected records is limited to QAO and DQAO to prevent unauthorized access or amendment.

Procedure for Records Management

- Identification: Records are uniquely identified.
- Collection: Observations, data and calculations are recorded at the time they are made. When mistakes are made in technical records, each mistake is crossed out with a single line (not erased, made illegible, or deleted) and the correct value entered alongside. Corrections are signed or initialed by the person making the correction. For electronic systems, all changes are tracked by the audit trail or by added notes. When changes are made to technical records for reasons other than for correction of transcription errors, the reason for the change is recorded on the document.
- Storage: All records stored on electronic media are supported by the hardware and software required for retrieval and have hard-copy or write-protected backup copies.
- Filing: Records are filed promptly and in an organized fashion.
- Access: Access to archived information is documented with an access log.
- Disposal: Records are disposed of according to applicable regulation, client request, or after seven years.

Backup/Disaster Recovery/Data Retention

The BRA IT Department performs backups of all local and cloud-based servers daily. In addition, local server backups are immediately copied to offsite cloud storage, and all cloud-based backups are kept for a total of ninety days, from which data or the entire server can be restored if needed. Instrument programs and electronic data are saved to the BRA network servers. In addition, local servers are replicated every ten to twenty minutes to cloud based disaster recovery and can be restored and made operable in the event of a total network failure at the main office.

Data Handling, Hardware, and Software Requirements

Water Quality Database (LIMS) - The Authority's laboratory database serves as a repository of water sample tracking and water quality analysis data until all appropriate tests and analyses have been performed and the results have undergone quality control review. The database resides on the Authority's network server, as described above, and is maintained through third party software application named Labvantage by Labvantage Solutions. Information Technology staff maintains the database which is installed on a Microsoft Virtual Server and is hosted on the Microsoft Azure Cloud, hosting the Labvantage Application as the front end and Microsoft SQL® as the back end. Data input and access to the laboratory water quality database are restricted by password and network access to the Laboratory Manager, Laboratory Staff, Field Operations Staff, QAO, DQAO, and the IT Software Administrator.

Information Resource Management Requirements

Data will be managed in accordance with the TCEQ DMRG (most recent revision) and applicable Brazos River Authority information resource management policies.

GPS equipment may be used as a component of the information required by the station location (SLOC) request process for creating the certified positional data that will ultimately be entered into SWQMIS database. Positional data obtained by CRP grantees using a GPS will follow the TCEQ's OPP 8.11 policy regarding the collection and management of positional data. Positional data may be acquired with a GPS and verified with photo interpolation using a certified source, such as Google Earth or Google Maps. The verified coordinates and map interface can then be used to develop a new SLOC.

C1 Assessments and Response Actions

The following table presents the types of assessments and response actions for data collection activities applicable to the QAPP.

Table C1.1 Assessments and Response Requirements

Assessment Activity	Approximate Schedule	Responsible Party	Scope	Response Requirements
Status Monitoring Oversight	Continuous	Brazos River Authority	Monitoring of the project status and records to ensure requirements are being fulfilled	Report to TCEQ in quarterly report. Submit CAPs to TCEQ as needed.
Monitoring Systems Audit of Brazos River Authority	Dates to be determined by TCEQ CRP	TCEQ	Field sampling, handling and measurement; facility review; and data management as they relate to CRP	30 days to provide corrective actions response to the TCEQ
Laboratory Assessment	Dates to be determined by TCEQ	TCEQ Laboratory Assessor	Analytical and quality control procedures employed at the laboratory and the contract laboratory	30 days to provide corrective actions response to the TCEQ

Corrective Action Process for Deficiencies

Deficiencies are any deviation from the QAPP, SWQM Procedures, DMRG, SOPs, or other applicable guidance documents. Deficiencies may invalidate resulting data and require corrective action. Deficiencies that can be prevented from occurring again in the future require a CAP. TCEQ QA staff recognize that deficiencies may occur that are out of the control of Brazos River Authority staff. Such deficiencies do not require a CAP.

However, when a deficiency impacts data quality or quantity, the TCEQ CRP PM must be notified (within three business days of discovery) and the data loss noted in the associated monitoring activities report and data summary. Corrective action for deficiencies may include for samples to be discarded and re-collected.

Deficiencies are documented in logbooks, field data sheets, etc. by field or laboratory staff, are communicated to the Brazos River Authority DQAO/QAO and PM (or other appropriate staff) and should be subject to periodic review so their responses can be uniform, and their frequency tracked. It is the responsibility of the Brazos River Authority PM, in consultation with the Brazos River Authority QAO, to ensure that the actions and resolutions to the problems are documented and that records are maintained in accordance with this QAPP.

TCEQ staff are tasked with reviewing CAPs written by Brazos River Authority concerning deficiencies associated with CRP work. This includes the TCEQ CRP Team Leader, PM, Project QAS, and Lead QAS. The Brazos River Authority PM or QAO should submit CAPs to their assigned TCEQ CRP PM in a timely manner. Brazos River Authority can begin implementing corrective actions without TCEQ approval. However, TCEQ may request alternate or modified corrective actions if deemed necessary.

A template for writing CAPs is provided in the [Guidance for Partners in the Texas Clean Rivers Program FY](#)

[2026–2027](#) (Exhibit 2C). While CAPs need not adhere to this specific format, they must include information for all of the listed elements. Incomplete CAPs will be returned to the Brazos River Authority QAO for revision. All CAPs for a FY should be cataloged in the quarterly progress reports submitted to the TCEQ CRP PM by the Brazos River Authority PM. This documentation should include, at a minimum, the report number, date(s) of deficiency occurrence, description of deficiency, action taken, CAP status, and the date the CAP was closed (if applicable).

Significant conditions that, if uncorrected, could have a serious effect on safety or on the validity or integrity of data will be reported to the TCEQ immediately.

The Brazos River Authority QAO is responsible for ensuring that corrective actions have been implemented and tracks deficiencies and corrective actions. Records of audit findings and corrective actions are maintained by the Brazos River Authority QAO. Audit reports and associated corrective action documentation will be submitted to the TCEQ with the quarterly progress reports.

If audit findings and corrective actions cannot be resolved, then the authority and responsibility for terminating work are specified in the TCEQ QMP and in agreements in contracts between participating organizations.

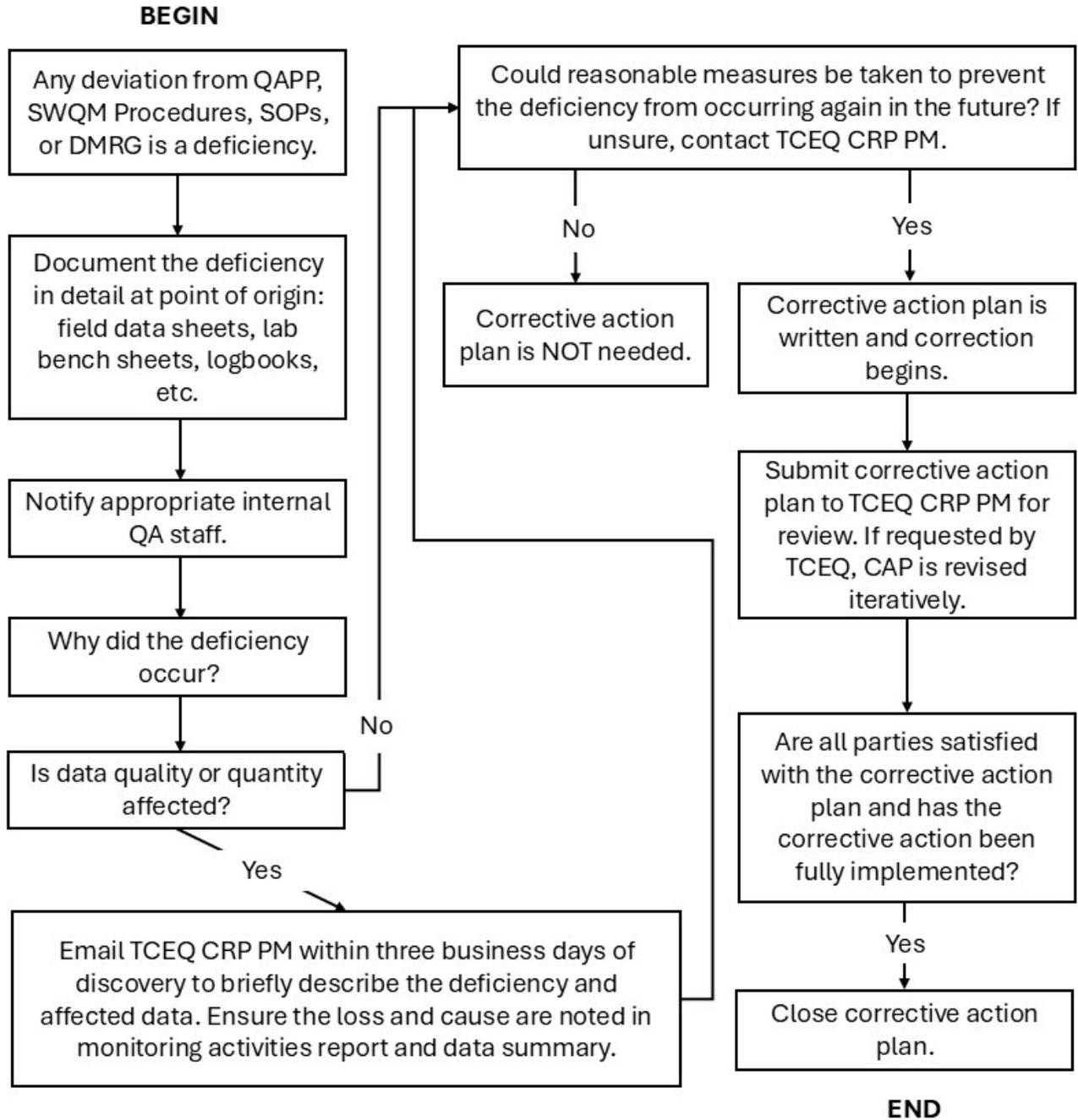
Corrective Action

CAPs should:

- Identify the problem, nonconformity, or undesirable situation
- Identify immediate remedial actions if possible
- Identify the underlying cause(s) of the problem
- Describe the programmatic impact
- Identify whether the problem is likely to recur, or occur in other areas
- Assist in determining the need for corrective action and actions to prevent reoccurrence
- Employ problem-solving techniques to verify causes, determine solution, and develop an action plan
- Identify personnel responsible for action
- Establish timelines and provide a schedule
- Document the corrective action and action(s) to prevent reoccurrence

A flow chart has been developed to facilitate the process (see Figure C1.1: Corrective Action Process for Deficiencies).

Figure C1.1 Corrective Action Process for Deficiencies



C2 Reports to Management

Table C2.1 QA Management Reports

Type of Report	Frequency (daily, weekly, monthly, quarterly, etc.)	Projected Delivery Date(s)	Person(s) Responsible for Report Preparation	Report Recipients
Corrective Action Plans	As Needed	As Needed	Field Staff Laboratory Staff	Brazos River Authority QA) and DQAO Brazos River Authority Laboratory Manager or Field Operations Manager as appropriate Brazos River Authority PM TCEQ CRP Project Manager
Data Completion Report	Monthly	One week following end of month	Brazos River Authority DQAO	Brazos River Authority PM
Internal Audit Reports	Quarterly	First week of following quarter	Brazos River Authority DQAO	Brazos River Authority PM Brazos River Authority QAO Brazos River Authority Laboratory Manager Brazos River Authority Field Operations Manager
Management Review	Annually	December of each year	Brazos River Authority QAO Brazos River Authority DQAO	Brazos River Authority Chief Environmental Officer Brazos River Authority PM Brazos River Authority Laboratory Manager Brazos River Authority Field Operations Manager
Progress Reports	Quarterly	December 15, 2025 March 15, 2026 June 15, 2026 September 15, 2026 December 15, 2026 March 15, 2027 June 15, 2027 August 15, 2027	Brazos River Authority PM	TCEQ CRP Project Manager
Monitoring Systems Audit Report and Response	As Needed	As Needed	Brazos River Authority QAO	TCEQ CRP Project Manager
Data Summary	Quarterly	December 1, 2025	Brazos River	TCEQ CRP Project

		March 1, 2026 May 1, 2026 August 1, 2026 December 1, 2026 March 1, 2027 May 1, 2027 August 1, 2027	Authority Data Manager	Manager
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Reports to Brazos River Authority Project Management

BRA does not anticipate reports from outside, non-BRA entities for the FY 26-27 biennium.

Corrective Action Report

Upon request from the BRA PM a Corrective Action Report is created once a Corrective Action has been found effective or not effective. The report includes the items found in C1 and can be printed or saved as pdf.

Data Completion Report

The QAO/DM or DQAO reports sampling, analysis, and QA activities to the BRA PM through a monthly data completion report. This report includes:

- Number of samples collected
- Number of samples analyzed per parameter
- Number of acceptable results per parameter
- Number of rejected results per parameter
- Completion percentage by month and year

Reports to TCEQ Project Management

All reports detailed in this section are contract deliverables and are transferred to the TCEQ in accordance with contract requirements.

Progress Report

Summarizes the Brazos River Authority’s activities for each task; reports monitoring status, problems, delays, deficiencies, status of open CAPs, and documentation for completed CAPs; and outlines the status of each task’s deliverables.

Monitoring Systems Audit Report and Response

Following any audit performed by the Brazos River Authority, a report of findings, recommendations and response is sent to the TCEQ in the quarterly progress report.

Data Summary

Contains basic identifying information about the data set and comments regarding inconsistencies and errors identified during data verification and validation steps or problems with data collection efforts (e.g., deficiencies).

Reports by TCEQ Project Management

Contractor Evaluation

The Brazos River Authority participates in a contractor evaluation by the TCEQ annually for compliance with administrative and programmatic standards. Results of the evaluation are submitted to the TCEQ Financial Administration Division, Procurement and Contracts Section.

D1 Data Review, Verification, and Validation

All field and laboratory data will be reviewed and verified for integrity, continuity, reasonableness, and conformance to project requirements, and then validated against the project objectives and measurement

performance specifications which are listed in Section A6 of this QAPP. Only those data which are supported by appropriate quality control data and meet the measurement performance specifications defined for this project will be considered acceptable and will be reported to the TCEQ for entry into SWQMIS.

Verification and Validation Methods

All field and laboratory data will be reviewed, verified and validated to ensure they conform to project specifications.

Data review, verification, and validation will be performed using self-assessments as well as peer and management review as appropriate to the project task. The data review tasks to be performed by field and laboratory staff are listed in the first two columns of Table D1.1. Potential errors are identified by examination of documentation and by manual examination of corollary or unreasonable data; this analysis may be computer-assisted. If a question arises or an error is identified, the manager of the task responsible for generating the data is contacted to resolve the issue. Issues which can be corrected are corrected and documented. If an issue cannot be corrected, the task manager consults with the higher-level project management to establish the appropriate course of action, or the data associated with the issue are rejected and not reported to the TCEQ for storage in SWQMIS. Field and laboratory reviews, verifications, and validations are documented.

After the field and laboratory data are reviewed, another level of review is performed once the data are combined into a data set. This review step, as specified in Table D1.1, is performed by the Brazos River Authority DM/QAO. Data review, verification, and validation tasks to be performed on the data set include, but are not limited to, the confirmation of laboratory and field data review, evaluation of field QC results, additional evaluation of anomalies and outliers, analysis of sampling and analytical gaps, and confirmation that all parameters and sampling sites are included in the QAPP.

The Data Review Checklist (see Appendix F) covers three main types of review: data format and structure, data quality review, and documentation review. The Data Review Checklist is completed and sent with the water quality data submitted to the TCEQ to ensure that the review process is being performed.

Another element of the data validation process is consideration of any findings identified during the monitoring systems audit conducted by the TCEQ CRP Lead QAS. Any issues requiring corrective action must be addressed, and the potential impact of these issues on previously collected data will be assessed. After the data are reviewed and documented, the Brazos River Authority PM validates that the data meet the data quality objectives of the project and are suitable for reporting to TCEQ.

If any requirements or specifications of the CRP are not met, based on any part of the data review, the responsible party should document the nonconforming activities and submit the information to the Brazos River Authority DM with the data in the data summary (See Appendix F). All failed QC checks, missing samples, missing analytes, missing parameters, and suspect results should be discussed in the data summary.

Table D1.1: Data Review Tasks

Data to be Verified	Field Task	Laboratory Task	QA Task	Lead Organization Data Manager Task
Sample documentation complete; samples labeled, sites identified	Field Staff		DQAO	
Standards and reagents traceable		Laboratory Analyst	DQAO	
Chain of custody complete/acceptable	Field Staff/Field Operations Manager	Laboratory Manager	DQAO	
NELAP Accreditation is current		Laboratory Analyst/Laboratory Manager	QAO/DQAO	
Sample preservation and handling acceptable	Field Staff	Laboratory Analyst	DQAO	
Holding times not exceeded	Field Staff	Laboratory Analyst	DQAO	
Collection, preparation, and analysis consistent with SOPs and QAPP	Field Staff/Field Operations Manager	Laboratory Analyst	DQAO	
Field documentation (e.g., biological, stream habitat) complete	Field Staff/Field Operations Manager		DQAO	
Instrument calibration data complete		Laboratory Analyst	DQAO	
QC samples analyzed at required frequency		Laboratory Analyst	DQAO	
QC results meet performance and program specifications		Laboratory Manager	DQAO	Data Manager
Analytical sensitivity (LOQ/AWRL) consistent with QAPP		Laboratory Manager	DQAO	Data Manager
Results, calculations, transcriptions checked		Laboratory Analyst/Laboratory Manager	DQAO	
Laboratory bench-level review performed		Laboratory Analyst/Laboratory Manager	DQAO	
All laboratory samples analyzed for all scheduled parameters		Laboratory Manager	DQAO	Data Manager
Corollary data agree			DQAO	Data Manager
Nonconforming activities documented	Field Staff/Field Operations Manager	Laboratory Analyst/Laboratory Manager	DQAO	Data Manager
Outliers confirmed and documented; reasonableness check performed			DQAO	Data Manager
Dates formatted correctly			DQAO	Data Manager

Depth reported correctly and in correct units	Field Staff		DQAO	Data Manager
TAG IDs correct			DQAO	Data Manager
TCEQ Station ID number assigned			DQAO	Data Manager
Valid parameter codes				Data Manager
Codes for submitting entity(ies), collecting entity(ies), and monitoring type(s) used correctly			DQAO	Data Manager
Time based on 24-hour clock			DQAO	Data Manager
Check for transcription errors				Data Manager
Sampling and analytical data gaps checked (e.g., all sites for which data are reported are on the coordinated monitoring schedule)			DQAO	Data Manager
Field instrument pre- and post-calibration check results within limits	Field Staff/Field Operations Manager		DQAO	
10% of data manually reviewed			DQAO	Data Manager

D2 Reconciliation with User Requirements

Data produced in this project, and data collected by other organizations (e.g., USGS, TCEQ, etc.), will be analyzed and reconciled with project data quality requirements. Data which do not meet requirements will not be submitted to SWQMIS nor will be considered appropriate for any of the uses noted in Section A4.

Appendix A: Measurement Performance Specifications (Table A6.1–9)

Measurement performance specifications define the data quality needed to satisfy project objectives. To this end, measurement performance specifications are qualitative and quantitative statements that:

- clarify the intended use of the data
- define the type of data needed to support the end use
- identify the conditions under which the data should be collected

Appendix A of the QAPP addresses measurement performance specifications, including:

- analytical methodologies
- AWRLs
- limits of quantitation
- bias limits for LCSs
- precision limits for laboratory control sample duplicates (LCSDs)
- completeness goals
- qualitative statements regarding representativeness and comparability

The items identified above should be considered for each type of monitoring activity. The CRP encourages that data be collected to address multiple objectives to optimize resources; however, caution should be applied when attempting to collect data for multiple purposes because measurement performance specifications may vary according to the purpose. For example, limits of quantitation may differ for data used to assess standards attainment and for trend analysis. When planning projects, first priority will be given to the main use of the project data and the data quality needed to support that use, then secondary goals will be considered.

Procedures for laboratory analysis must be in accordance with the most recently published edition of Standard Methods for the Examination of Water and Wastewater, 40 CFR 136, or otherwise approved independently. Only data collected that have a valid TCEQ parameter code assigned in Tables A6 are stored in SWQMIS. Any parameters listed in Tables A6 that do not have a valid TCEQ parameter code assigned will not be stored in SWQMIS.

Table A6.1–9 : Measurement Performance Specifications

TABLE A6.1 Measurement Performance Specifications for Field Parameters					
Field Parameters					
Parameter	Units	Matrix	Method	Parameter Code	Lab
TEMPERATURE, WATER (DEGREES CENTIGRADE)	DEG C	water	SM 2550 B and TCEQ SOP V1	00010	Field
TRANSPARENCY, SECCHI DISC (METERS)	meters	water	TCEQ SOP V1	00078	Field
SPECIFIC CONDUCTANCE, FIELD (US/CM @ 25C)	us/cm	water	EPA 120.1 and TCEQ SOP, V1	00094	Field
OXYGEN, DISSOLVED (MG/L)	mg/L	water	SM 4500-O G and TCEQ SOP V1	00300	Field
PH (STANDARD UNITS)	s.u.	water	EPA 150.1 and TCEQ SOP V1	00400	Field
DAYS SINCE PRECIPITATION EVENT (DAYS)	days	other	TCEQ SOP V1	72053	Field
RESERVOIR STAGE (FEET ABOVE MEAN SEA LEVEL)***	FT ABOVE MSL	water	TWDB	00052	Field
RESERVOIR PERCENT FULL***	% RESERVOIR CAPACITY	water	TWDB	00053	Field
RESERVOIR ACCESS NOT POSSIBLE LEVEL TOO LOW ENTER 1 IF REPORTING	NS	other	TCEQ Drought Guidance	00051	Field
AVERAGE STREAM WIDTH	meters	other	TCEQ SOP V2	89861	Field
MAXIMUM POOL WIDTH AT TIME OF STUDY (METERS)**	meters	other	TCEQ SOP V2	89864	Field
MAXIMUM POOL DEPTH AT TIME OF STUDY (METERS)**	meters	other	TCEQ SOP V2	89865	Field
POOL LENGTH, METERS**	meters	other	TCEQ SOP V2	89869	Field
% POOL COVERAGE IN 500 METER REACH**	%	other	TCEQ SOP V2	89870	Field
MACROPHYTE BED AT COLLECTION POINT	%	other	TCEQ SOP V2	89926	Field
WIND INTENSITY (1=CALM,2=SLIGHT,3=MOD.,4=STRONG)	NU	other	NA	89965	Field
PRESENT WEATHER (1=CLEAR,2=PTCLDY,3=CLDY,4=RAIN,5=OTHER)	NU	other	NA	89966	Field
WATER SURFACE (1=CALM,2=RIPPLE,3=WAVE,4=WHITECAP)	NU	water	NA	89968	Field
<p>* Reporting to be consistent with SWQM guidance and based on measurement capability. ** To be routinely reported when collecting data from perennial pools. *** As published by the Texas Water Development Board on their website https://www.waterdatafortexas.org/reservoirs/statewide</p> <p>References: United States Environmental Protection Agency (USEPA), Clean Water Act Analytical Methods Standard Methods for the Examination of Water and Wastewater, 24th Edition, 2022 or applicable version TCEQ SOP, V1 - TCEQ Surface Water Quality Monitoring Procedures, Volume 1: Physical and Chemical Monitoring Methods, 2012 (RG-415). TCEQ SOP, V2 - TCEQ Surface Water Quality Monitoring Procedures, Volume 2: Methods for Collecting and Analyzing Biological Assemblage and Habitat Data, 2014 (RG-416).</p>					

TABLE A6.2 Measurement Performance Specifications for Flow Parameters

Flow Parameters					
Parameter	Units	Matrix	Method	Parameter Code	Lab
FLOW STREAM, INSTANTANEOUS (CUBIC FEET PER SEC)	cfs	water	TCEQ SOP V1	00061	Field
FLOW SEVERITY:1=No Flow,2=Low,3=Normal,4=Flood,5=High,6=Dry	NU	water	TCEQ SOP V1	01351	Field
FLOW MTH 1=GAGE 2=ELEC 3=MECH 4=WEIR/FLU 5=DOPPLER	NU	other	TCEQ SOP V1	89835	Field
References: TCEQ SOP, V1 - TCEQ Surface Water Quality Monitoring Procedures, Volume 1: Physical and Chemical Monitoring Methods, 2012 (RG-415).					

TABLE A6.3 Measurement Performance Specifications for Conventional Parameters in Water

Conventional Parameters in Water										
Parameter	Units	Matrix	Method	Parameter Code	TCEQ AWRL	LOQ	LOQ Check Sample %Rec*	Precision (RPD)	Bias %Rec. of LCS	Lab
RESIDUE, TOTAL NONFILTRABLE (MG/L)	mg/L	water	SM 2540D	00530	5	5	NA	10	80-120	BRA
NITROGEN, AMMONIA, TOTAL (MG/L AS N)	mg/L	water	EPA 350.1 Rev. 2.0 (1993)	00610	0.1	0.05	70-130	20	80-120	BRA
NITRITE NITROGEN, TOTAL (MG/L AS N)	mg/L	water	EPA 300.0 Rev. 2.1 (1993)	00615	0.05	0.04	70-130	20	80-120	BRA
NITRATE NITROGEN, TOTAL (MG/L AS N)	mg/L	water	EPA 300.0 Rev. 2.1 (1993)	00620	0.05	0.04	70-130	20	80-120	BRA
NITROGEN, KJELDAHL, TOTAL (MG/L AS N)	mg/L	water	EPA 351.2 Rev. 2.0	00625	0.2	0.2	70-130	20	80-120	BRA
PHOSPHORUS, TOTAL, WET METHOD (MG/L AS P)	mg/L	water	EPA 365.1 Rev 2.0**	00665	0.06	0.02	70-130	20	80-120	BRA
ORTHOPHOSPHATE PHOSPHORUS, DISS, MG/L, FLDFILT < 15MIN	mg/L	water	EPA 300.0 Rev. 2.1 (1993)	00671	0.04	0.04	70-130	20	80-120	BRA
CHLORIDE (MG/L AS CL)	mg/L	water	EPA 300.0 Rev. 2.1 (1993)	00940	5	5	70-130	20	80-120	BRA
SULFATE (MG/L AS SO4)	mg/L	water	EPA 300.0 Rev. 2.1 (1993)	00945	5	5	70-130	20	80-120	BRA
RESIDUE, TOT DISS, UNSPEC CALC BASED ON COND (MG/	mg/L	water	calculation	70294**	NA	NA	NA	NA	NA	BRA
RESIDUE, TOTAL FILTRABLE (DRIED AT 180C) (MG/L)	mg/L	water	SM 2540C	70300	10	10	NA	10	80-120	BRA
ORTHOPHOSPHATE PHOSPHORUS, DISS, MG/L, FILTER >15MIN	mg/L	water	EPA 300.0 Rev. 2.1 (1993)	70507***	0.04	0.04	70-130	20	80-120	BRA
CHLOROPHYLL-A, FLUOROMETRIC METHOD, UG/L	µg/L	water	EPA 445.0 Rev. 1.2	70953	3	3	NA	20	80-120	BRA
TURBIDITY, LAB NEPHELOMETRIC TURBIDITY UNITS, NTU	NTU	water	SM 2130B	82079	0.5	0.5	70-130	20	80-120	BRA

*When the acceptance criteria are exceeded (high bias) and there are associated samples that are <LOQ, those samples may be reported.
 **Currently subcontracted to Aqua-Tech Laboratories until BRA acquires accreditation. Paperwork for accreditation will be submitted in August 2025.
 ***This parameter is only reported under special circumstances when 70300 cannot be reported.
 ****This parameter is only reported under special circumstances when 00671 cannot be reported.

References:
 United States Environmental Protection Agency (USEPA), Clean Water Act Analytical Methods
 Standard Methods for the Examination of Water and Wastewater, 24th Edition, 2022 or applicable version
 TCEQ SOP, V1 - TCEQ Surface Water Quality Monitoring Procedures, Volume 1: Physical and Chemical Monitoring Methods, 2012 (RG-415).

TABLE A6.4 Measurement Performance Specifications for Bacteriological Parameters in Water

Bacteriological Parameters in Water										
Parameter	Units	Matrix	Method	Parameter Code	TCEQ AWRL	LOQ	LOQ Check Sample %Rec	Log Difference of Duplicates	Bias %Rec. of LCS	Lab
E. COLI, COLILERT, IDEXX METHOD, MPN/100ML	MPN/100 mL	water	IDEXX Laboratories Colilert©**	31699	1	1	NA	0.50*	NA	BRA
E. COLI, COLILERT, IDEXX METHOD, MPN/100ML	MPN/100 mL	water	IDEXX Laboratories Colilert©-18**	31699	1	1(10)***	NA	0.50*	NA	BRA
ENTEROCOCCI, ENTEROLERT, IDEXX, (MPN/100 ML)	MPN/100 mL	water	IDEXX Laboratories Enterolert©	31701	10***	10	NA	0.50*	NA	BRA
E.COLI, COLILERT, IDEXX, HOLDING TIME	hours	water	NA	31704	NA	NA	NA	NA	NA	BRA

* This value is not expressed as a relative percent difference. It represents the maximum allowable difference between the logarithm of the result of a sample and the logarithm of the duplicate result. See Section B4.

** *E.coli* samples analyzed by these methods should always be processed as soon as possible and within 8 hours. When transport conditions necessitate delays in delivery longer than 6 hours, the holding time may be extended and samples must be processed as soon as possible and within 30 hours.

*** *Enterococcus* samples should be diluted 1:10 for all waters. *E. coli* samples with specific conductance greater than 3000 uS/cm should be diluted 1:10 using Colilert 18 media.

References:

Standard Methods for the Examination of Water and Wastewater, 24th Edition, 2022 or applicable version

Annual Book of ASTM Standards, Section 11, Water and Environmental Technology, Volume 11.02, Water

TCEQ SOP, V1 - TCEQ Surface Water Quality Monitoring Procedures, Volume 1: Physical and Chemical Monitoring Methods, 2012 (RG-415).

TABLE A6.5 Measurement Performance Specifications for 24-hr W Parameters

24 Hour Parameters in Water					
Parameter	Units	Matrix	Method	Parameter Code	Lab
TEMPERATURE, WATER (DEGREES CENTIGRADE), 24HR AVG	DEG C	Water	TCEQ SOP V1	00209	Field
WATER TEMPERATURE, DEGREES CENTIGRADE, 24HR MAX	DEG C	Water	TCEQ SOP V1	00210	Field
TEMPERATURE, WATER (DEGREES CENTIGRADE) 24HR MIN	DEG C	Water	TCEQ SOP V1	00211	Field
SPECIFIC CONDUCTANCE, US/CM, FIELD, 24HR AVG	uS/cm	Water	TCEQ SOP V1	00212	Field
SPECIFIC CONDUCTANCE, US/CM, FIELD, 24HR MAX	uS/cm	Water	TCEQ SOP V1	00213	Field
SPECIFIC CONDUCTANCE, US/CM, FIELD, 24HR MIN	uS/cm	Water	TCEQ SOP V1	00214	Field
PH, S.U., 24HR MAXIMUM VALUE	std. units	Water	TCEQ SOP V1	00215	Field
PH, S.U., 24HR, MINIMUM VALUE	std. units	Water	TCEQ SOP V1	00216	Field
SALINITY, 24-HR, MAXIMUM, PPT	ppt	Water	TCEQ SOP V1	00217	Field
SALINITY, 24-HR, AVERAGE, PPT	ppt	Water	TCEQ SOP V1	00218	Field
SALINITY, 24-HR, MINIMUM, PPT	ppt	Water	TCEQ SOP V1	00219	Field
SALINITY, # OF MEASUREMENTS IN 24-HRS	NU	Water	TCEQ SOP V1	00220	Field
WATER TEMPERATURE, # OF MEASUREMENTS IN 24-HRS	NU	Water	TCEQ SOP V1	00221	Field
SPECIFIC CONDUCTANCE, # OF MEASUREMENTS IN 24-HRS	NU	Water	TCEQ SOP V1	00222	Field
pH, # OF MEASUREMENTS IN 24-HRS	NU	Water	TCEQ SOP V1	00223	Field
DISSOLVED OXYGEN, 24-HOUR MIN. (MG/L) MIN. 4 MEA	mg/l	Water	TCEQ SOP V1	89855	Field
DISSOLVED OXYGEN, 24-HOUR MAX. (MG/L) MIN. 4 MEA	mg/l	Water	TCEQ SOP V1	89856	Field
DISSOLVED OXYGEN, 24-HOUR AVG. (MG/L) MIN. 4 MEA	mg/l	Water	TCEQ SOP V1	89857	Field
DISSOLVED OXYGEN, # OF MEASUREMENTS IN 24-HRS	NU	Water	TCEQ SOP V1	89858	Field
References: TCEQ SOP, V1 - TCEQ Surface Water Quality Monitoring Procedures, Volume 1: Physical and Chemical Monitoring Methods, 2012 (RG-415).					

TABLE A6.6 Measurement Performance Specifications for Bio Habitat Parameters

Biological - Habitat					
Parameter	Units	Matrix	Method	Parameter Code	Lab
FLOW STREAM, INSTANTANEOUS (CUBIC FEET PER SEC)	cfs	Water	TCEQ SOP V2	00061	Field
BIOLOGICAL DATA	NS	Other	NA/Calculation	89888	Field
STREAM TYPE; 1=PERENNIAL 2=INTERMITTENT S/PERENNIAL POOLS 3=INTERMITTENT 4=UNKNOWN	NU	Water	NA/Calculation	89821	Field
STREAMBED SLOPE (M/KM)	M/KM	Other	NA/Calculation	72051	Field
ECOREGION LEVEL III (TEXAS ECOREGION CODE)	NU	Other	TCEQ SOP V1	89961	Field
AVERAGE PERCENTAGE INSTREAM COVER	%	Other	TCEQ SOP V2	84159	Field
STREAM ORDER	NU	Water	TCEQ SOP V2	84161	Field
NUMBER OF LATERAL TRANSECTS MADE	NU	Other	TCEQ SOP V2	89832	Field
FLOW MTH 1=GAGE 2=ELEC 3=MECH 4=WEIR/FLU 5=DOPPLER	NU	Other	TCEQ SOP V2	89835	Field
TOTAL NUMBER OF STREAM BENDS	NU	Other	TCEQ SOP V2	89839	Field
NUMBER OF WELL DEFINED STREAM BENDS	NU	Other	TCEQ SOP V2	89840	Field
NUMBER OF MODERATELY DEFINED STREAM BENDS	NU	Other	TCEQ SOP V2	89841	Field
NUMBER OF POORLY DEFINED STREAM BENDS	NU	Other	TCEQ SOP V2	89842	Field
TOTAL NUMBER OF RIFFLES	NU	Other	TCEQ SOP V2	89843	Field
DOMINANT SUBSTRATE TYPE(1=CLAY,2=SILT,3=SAND,4=GRAVEL,5=COBBLE,6=BOULDER,7=BEDROCK,8=OTHER)	NU	Sediment	TCEQ SOP V2	89844	Field
AVERAGE PERCENT OF SUBSTRATE GRAVEL SIZE OR LARGER	%	Other	TCEQ SOP V2	89845	Field
AVERAGE STREAM BANK EROSION (%)	%	Other	TCEQ SOP V2	89846	Field
AVERAGE STREAM BANK SLOPE (DEGREES)	deg	Other	TCEQ SOP V2	89847	Field
HABITAT FLOW STATUS, 1=NO FLOW, 2=LOW,3=MOD,4=HIGH	NU	Other	TCEQ SOP V2	89848	Field
AVERAGE PERCENT TREES AS RIPARIAN VEGETATION	%	Other	TCEQ SOP V2	89849	Field
AVERAGE PERCENT SHRUBS AS RIPARIAN VEGETATION	%	Other	TCEQ SOP V2	89850	Field
AVERAGE PERCENT GRASS AS RIPARIAN VEGETATION	%	Other	TCEQ SOP V2	89851	Field
AVERAGE PERCENT CULTIVATED FIELDS AS RIPARIAN VEGETATION	%	Other	TCEQ SOP V2	89852	Field
AVERAGE PERCENT OTHER AS RIPARIAN VEGETATION	%	Other	TCEQ SOP V2	89853	Field
AVERAGE PERCENTAGE OF TREE CANOPY COVERAGE	%	Other	TCEQ SOP V2	89854	Field
DRAINAGE AREA ABOVE MOST DOWNSTREAM TRANSECT*	km2	Other	TCEQ SOP V2	89859	Field
REACH LENGTH OF STREAM EVALUATED (M)	m	Other	NA/Calculation	89884	Field
AVERAGE STREAM WIDTH (METERS)	M	Other	TCEQ SOP V2	89861	Field
AVERAGE STREAM DEPTH (METERS)	M	Other	TCEQ SOP V2	89862	Field
MAXIMUM POOL WIDTH AT TIME OF STUDY (METERS)	M	Other	TCEQ SOP V2	89864	Field
MAXIMUM POOL DEPTH AT TIME OF STUDY(METERS)	M	Other	TCEQ SOP V2	89865	Field
AVERAGE WIDTH OF NATURAL RIPARIAN VEGETATION (M)	M	Other	TCEQ SOP V2	89866	Field
AVERAGE WIDTH OF NATURAL RIPARIAN BUFFER ON LEFT BANK (M)	M	Other	NA/Calculation	89872	Field
AVERAGE WIDTH OF NATURAL RIPARIAN BUFFER ON RIGHT BANK (M)	m	Other	NA/Calculation	89873	Field

AESTHETICS OF REACH (1=WILD 2=NAT. 3=COMM. 4=OFF.)	NU	Other	TCEQ SOP V2	89867	Field
NUMBER OF STREAM COVER TYPES	NU	Other	TCEQ SOP V2	89929	Field
LAND DEVELOP IMPACT (1=UNIMP,2=LOW,3=MOD,4=HIGH)	NU	Other	TCEQ SOP V2	89962	Field
RIPARIAN VEGETATION %; LEFT BANK - TREES	%	Other	NA/Calculation	89822	Field
RIPARIAN VEGETATION %; RIGHT BANK - TREES	%	Other	NA/Calculation	89823	Field
RIPARIAN VEGETATION %; LEFT BANK SHRUBS	%	Other	NA/Calculation	89824	Field
RIPARIAN VEGETATION %; RIGHT BANK - SHRUBS	%	Other	NA/Calculation	89825	Field
RIPARIAN VEGETATION %: LEFT BANK - GRASSES OR FORBS	%	Other	NA/Calculation	89826	Field
RIPARIAN VEGETATION %; RIGHT BANK - GRASSES OR FORBS	%	Other	NA/Calculation	89827	Field
RIPARIAN VEGETATION %: LEFT BANK - CULTIVATED FIELDS	%	Other	NA/Calculation	89828	Field
RIPARIAN VEGETATION %: RIGHT BANK - CULTIVATED FIELDS	%	Other	NA/Calculation	89829	Field
RIPARIAN VEGETATION %: LEFT BANK - OTHER	%	Other	NA/Calculation	89830	Field
RIPARIAN VEGETATION %: RIGHT BANK - OTHER	%	Other	NA/Calculation	89871	Field
AVAILABLE INSTREAM COVER HQI SCORE: 4=ABUNDANT 3=COMMON 2=RARE 1=ABSENT	NU	Other	NA/Calculation	89874	Field
BOTTOM SUBSTRATE STABILITY HQI SCORE: 4=STABLE 3=MODERATELY STABLE 2=MODERATELY UNSTABLE 1=UNSTABLE	NU	Other	NA/Calculation	89875	Field
NUMBER OF RIFFLES HQI SCORE: 4=ABUNDANT 3=COMMON 2=RARE 1=ABSENT	NS	Other	NA/Calculation	89876	Field
DIMENSIONS OF LARGEST POOL HQI SCORE: 4=LARGE 3=MODERATE 2=SMALL 1=ABSENT	NU	Other	NA/Calculation	89877	Field
CHANNEL FLOW STATUS HQI SCORE: 3=HIGH 2=MODERATE 1=LOW 0=NO FLOW	NU	Other	NA/Calculation	89878	Field
BANK STABILITY HQI SCORE: 3=STABLE 2=MODERATELY STABLE 1=MODERATELY UNSTABLE 0=UNSTABLE	NU	Other	NA/Calculation	89879	Field
CHANNEL SINUOSITY HQI SCORE: 3=HIGH 2=MODERATE 1=LOW 0=NONE	NU	Other	NA/Calculation	89880	Field
RIPARIAN BUFFER VEGETATION HQI SCORE: 3=EXTENSIVE 2=WIDE 1=MODERATE 0=NARROW	NU	Other	NA/Calculation	89881	Field
AESTHETICS OF REACH HQI SCORE: 3=WILDERNESS 2=NATURAL AREA 1=COMMON SETTING 0=OFFENSIVE	NU	Other	NA/Calculation	89882	Field
HQI TOTAL SCORE	NU	Other	NA/Calculation	89883	Field
NO FLOW ISOLATED POOL: LARGEST POOL MAX WIDTH (M)	M	Other	NA/Calculation	89908	Field
NO FLOW ISOLATED POOL: LARGEST POOL MAX LENGTH (M)	M	Other	NA/Calculation	89909	Field
NO FLOW ISOLATED POOL: LARGEST POOL MAX DEPTH (M)	M	Other	NA/Calculation	89910	Field
NO FLOW ISOLATED POOL: SMALLEST POOL MAX DEPTH (M)	M	Other	NA/Calculation	89911	Field
NO FLOW ISOLATED POOL: SMALLEST POOL MAX WIDTH (M)	M	Other	NA/Calculation	89912	Field
NO FLOW ISOLATED POOL: SMALLEST POOL MAX LENGTH (M)	M	Other	NA/Calculation	89913	Field
NO FLOW ISOLATED POOLS: NUMBER OF POOLS EVALUATED	NU	Other	NA/Calculation	89914	Field

* From USGS map.

References:

TCEQ SOP, V1 - TCEQ Surface Water Quality Monitoring Procedures, Volume 1: Physical and Chemical Monitoring Methods, 2012 (RG-415).

TCEQ SOP, V2 - TCEQ Surface Water Quality Monitoring Procedures, Volume 2: Methods for Collecting and Analyzing Biological Assemblage and Habitat Data, 2014 (RG-416).

TABLE A6.7 Measurement Performance Specifications for Biological Benthic Parameters

Biological - Benthics (Quantitative)					
Parameter	Units	Matrix	Method	Parameter Code	Lab
AREA OF SNAG SURFACE SAMPLED (SQ.MT)	m2	Other	TCEQ SOP V2	89975	Field
BENTHIC GRAZERS, PERCENT OF INDIVIDUALS	%	Other	TCEQ SOP V2	90020	Field
BENTHIC GATHERERS, PERCENT OF INDIVIDUALS	%	Other	TCEQ SOP V2	90025	Field
BENTHIC FILTERERS, PERCENT OF INDIVIDUALS	%	Other	TCEQ SOP V2	90030	Field
NUMBER OF EPHEMEROPTERA TAXA	NU	Other	TCEQ SOP V2	90057	Field
TOTAL NUMBER OF INTOLERANT TAXA, BENTHOS	NU	Other	TCEQ SOP V2	90058	Field
EPT, PERCENT OF INDIVIDUALS	%	Other	TCEQ SOP V2	90060	Field
TOLERANT BENTHOS, PERCENT OF INDIVIDUALS	%	Other	TCEQ SOP V2	90066	Field

References:

TCEQ SOP, V2 - TCEQ Surface Water Quality Monitoring Procedures, Volume 2: Methods for Collecting and Analyzing Biological Assemblage and Habitat Data, 2014 (RG-416).

TABLE A6.8 Measurement Performance Specifications for Biological Benthic Parameters

Biological - Benthics (Qualitative)					
Parameter	Units	Matrix	Method	Parameter Code	Lab
STREAM ORDER	NU	Water	TCEQ SOP, V1	84161	Field
BIOLOGICAL DATA	NS	Other	NA/Calculation	89888	Field
RAPID BIOASSESSMENT PROTOCOLS BENTHIC MACROINVERTEBRATE IBI SCORE	NS	Other	NA/Calculation	90081	Field
BENTHIC DATA REPORTING UNITS (1=NUMBER OF INDIVIDUALS IN SUB-SAMPLE, 2=NUMBER OF INDIVIDUALS/FT2, 3=NUMBER OF INDIVIDUALS/M2, 4=TOTAL NUMBER OF INDIVIDUALS IN SAMPLE)	NU	Other	TCEQ SOP V2	89899	Field
KICKNET EFFORT,AREA KICKED (SQ.METER)	m2	Other	TCEQ SOP V2	89903	Field
KICKNET EFFORT,MINUTES KICKED (MIN.)	min.	Other	TCEQ SOP V2	89904	Field
DEBRIS/SHORELINE SAMPLING EFFORT, MINUTES	min.	Other	TCEQ SOP V2	89905	Field
NUMBER OF INDIVIDUALS IN BENTHIC SAMPLE	NU	Other	TCEQ SOP V2	89906	Field
UNDERCUT BANK AT COLLECTION POINT (%)	%	Other	TCEQ SOP V2	89921	Field
OVERHANGING BRUSH AT COLLECTION POINT (%)	%	Other	TCEQ SOP V2	89922	Field
GRAVEL BOTTOM AT COLLECTION POINT (%)	%	Sediment	TCEQ SOP V2	89923	Field
SAND BOTTOM AT COLLECTION POINT (%)	%	Sediment	TCEQ SOP V2	89924	Field
SOFT BOTTOM AT COLLECTION POINT (%)	%	Sediment	TCEQ SOP V2	89925	Field
MACROPHYTE BED AT COLLECTION POINT (%)	%	Other	TCEQ SOP V2	89926	Field
SNAGS AND BRUSH AT COLLECTION POINT (%)	%	Other	TCEQ SOP V2	89927	Field
BEDROCK STREAMBED AT COLLECTION POINT (%)	%	Sediment	TCEQ SOP V2	89928	Field
MESH SIZE, ANY NET OR SIEVE, AVERAGE BAR (CM)	cm	Other	TCEQ SOP V2	89946	Field
BENTHIC SAMPLE COLLECTION METHOD (1=SURBER, 2=EKMAN, 3=KICKNET, 4=PETERSON, 5=HESTER DENDY, 6=SNAG, 7=HESS)	NU	Other	TCEQ SOP V2	89950	Field
ECOREGION LEVEL III (TEXAS ECOREGION CODE)	NU	Other	TCEQ SOP V1	89961	Field
BENTHOS ORGANISMS -NONE PRESENT (0=NONE PRESENT)	NS	Other	TCEQ SOP V2	90005	Field
HILSENHOFF BIOTIC INDEX (HBI)	NU	Other	TCEQ SOP V2	90007	Field
NUMBER OF EPT INDEX	NU	Other	TCEQ SOP V2	90008	Field
DOMINANT BENTHIC FUNCTIONAL FEEDING GRP, % OF INDIVIDUALS	%	Other	TCEQ SOP V2	90010	Field
BENTHIC GATHERERS, PERCENT OF INDIVIDUALS	%	Other	TCEQ SOP V2	90025	Field
BENTHIC PREDATORS, PERCENT OF INDIVIDUALS	%	Other	TCEQ SOP V2	90036	Field
DOMINANT TAXON, BENTHOS PERCENT OF INDIVIDUALS	%	Other	TCEQ SOP V2	90042	Field
RATIO OF INTOLERANT TO TOLERANT TAXA, BENTHOS	NU	Other	TCEQ SOP V2	90050	Field
NUMBER OF NON-INSECT TAXA	NU	Other	TCEQ SOP V2	90052	Field
ELMIDAE, PERCENT OF INDIVIDUALS	%	Other	TCEQ SOP V2	90054	Field
TOTAL TAXA RICHNESS, BENTHOS	NU	Other	TCEQ SOP V2	90055	Field
CHIRONOMIDAE, PERCENT OF INDIVIDUALS	%	Other	TCEQ SOP V2	90062	Field
PERCENT OF TOTAL TRICHOPTERA INDIVIDUALS AS HYDROPSYCHIDAE	%	Other	TCEQ SOP V2	90069	Field
TOTAL # OF BENTHIC GENERA IN SAMPLE	NU	Other	TCEQ SOP V2	90011	Field

TABLE A6.8 Measurement Performance Specifications for Biological Benthic Parameters

Biological - Benthics (Qualitative)					
Parameter	Units	Matrix	Method	Parameter Code	Lab
BENTHIC SHREDDERS (% OF COMMUNITY)	%	Other	TCEQ SOP V2	90035	Field
TOTAL # OF FAMILIES IN BENTHIC SAMPLE	NU	Other	TCEQ SOP V2	90012	Field
References: TCEQ SOP, V1 - TCEQ Surface Water Quality Monitoring Procedures, Volume 1: Physical and Chemical Monitoring Methods, 2012 (RG-415). TCEQ SOP, V2 - TCEQ Surface Water Quality Monitoring Procedures, Volume 2: Methods for Collecting and Analyzing Biological Assemblage and Habitat Data, 2014 (RG-416).					

TABLE A6.9 Measurement Performance Specifications for Bio Nekton Parameters

Biological - Nekton					
Parameter	Units	Matrix	Method	Parameter Code	Lab
STREAM ORDER	NU	Water	TCEQ SOP V1	84161	Field
NEKTON TEXAS REGIONAL IBI SCORE	NS	Other	NA/Calculation	98123	Field
BIOLOGICAL DATA	NS	Other	NA/Calculation	89888	Field
SEINE, MINIMUM MESH SIZE, AVERAGE BAR, NEKTON,IN	IN	Other	TCEQ SOP V2	89930	Field
SEINE, MAXIMUM MESH SIZE, AVG BAR, NEKTON,INCH	IN	Other	TCEQ SOP V2	89931	Field
NET LENGTH (METERS)	M	Other	TCEQ SOP V2	89941	Field
ELECTROFISHING METHOD 1=BOAT 2=BACKPACK 3=TOTEBARGE	NU	Other	TCEQ SOP V2	89943	Field
ELECTROFISH EFFORT, DURATION OF SHOCKING (SEC)	SEC	Other	TCEQ SOP V2	89944	Field
SEINING EFFORT (# OF SEINE HAULS)	NU	Other	TCEQ SOP V2	89947	Field
COMBINED LENGTH OF SEINE HAULS (METERS)	M	Other	TCEQ SOP V2	89948	Field
SEINING EFFORT, DURATION (MINUTES)	MIN	Other	TCEQ SOP V2	89949	Field
ECOREGION LEVEL III (TEXAS ECOREGION CODE)	NU	Other	TCEQ SOP V1	89961	Field
AREA SEINED (SQ METERS)	M2	Other	TCEQ SOP V2	89976	Field
NUMBER OF SPECIES, FISH	NU	Other	TCEQ SOP V2	98003	Field
NEKTON ORGANISMS-NONE PRESENT (0=NONE PRESENT)	NS	Other	TCEQ SOP V2	98005	Field
TOTAL NUMBER OF SUNFISH SPECIES	NU	Other	TCEQ SOP V2	98008	Field
TOTAL NUMBER OF INTOLERANT SPECIES, FISH	NU	Other	TCEQ SOP V2	98010	Field
PERCENT OF INDIVIDUALS AS OMNIVORES, FISH	%	Other	TCEQ SOP V2	98017	Field
PERCENT OF INDIVIDUALS AS INVERTIVORES, FISH	%	Other	TCEQ SOP V2	98021	Field
PERCENT OF INDIVIDUALS AS PISCIVORES, FISH	%	Other	TCEQ SOP V2	98022	Field
PERCENT OF INDIVIDUALS WITH DISEASE OR ANOMALY	%	Other	TCEQ SOP V2	98030	Field
TOTAL NUMBER OF NATIVE CYPRINID SPECIES	NU	Other	TCEQ SOP V2	98032	Field
PERCENT INDIVIDUALS AS NON-NATIVE FISH SPECIES (% OF COMMUNITY)	%	Other	TCEQ SOP V2	98033	Field
TOTAL NUMBER OF INDIVIDUALS SEINING	NU	Other	TCEQ SOP V2	98039	Field
TOTAL NUMBER OF INDIVIDUALS ELECTROFISHING	NU	Other	TCEQ SOP V2	98040	Field
TOTAL NUMBER OF BENTHIC INVERTIVORE SPECIES	NU	Other	TCEQ SOP V2	98052	Field
TOTAL NUMBER OF BENTHIC FISH SPECIES	NU	Other	TCEQ SOP V2	98053	Field
NUMBER OF INDIVIDUALS PER SEINE HAUL	NU	Other	TCEQ SOP V2	98062	Field
NUMBER OF INDIVIDUALS PER MINUTE ELECTROFISHING	NU	Other	TCEQ SOP V2	98069	Field
PERCENT INDIVIDUALS AS TOLERANT FISH SPECIES (EXCLUDING WESTERN MOSQUITOFISH)	%	Other	TCEQ SOP V2	98070	Field
TOTAL NUMBER OF SUCKER SPECIES	NU	Other	TCEQ SOP V2	98009	Field
PERCENT OF INDIVIDUALS AS HYBRIDS	%	Other	TCEQ SOP V2	98024	Field
TOTAL NUMBER OF INDIVIDUALS IN SAMPLE, FISH	NU	Other	TCEQ SOP V2	98023	Field

TABLE A6.9 Measurement Performance Specifications for Bio Nekton Parameters

Biological - Nekton					
Parameter	Units	Matrix	Method	Parameter Code	Lab
PERCENT OF INDIVIDUALS AS TOLERANTS, FISH	%	Other	TCEQ SOP V2	98016	Field
TOTAL NUMBER OF DARTER SPECIES	NU	Other	TCEQ SOP V2	98004	Field
References: TCEQ SOP, V1 - TCEQ Surface Water Quality Monitoring Procedures, Volume 1: Physical and Chemical Monitoring Methods, 2012 (RG-415). TCEQ SOP, V2 - TCEQ Surface Water Quality Monitoring Procedures, Volume 2: Methods for Collecting and Analyzing Biological Assemblage and Habitat Data, 2014 (RG-416).					

Appendix B: Task 3 Work Plan & Sampling Process Design and Monitoring Schedule (Plan)

Objective: Water quality monitoring will focus on the characterization of a variety of locations and conditions. This will include a combination of the following:

- Planning and coordinating basin-wide monitoring.
- Routine, regularly scheduled monitoring to collect long-term information and support statewide assessment of water quality.
- Systematic, regularly scheduled short-term monitoring to screen water bodies for issues.

Task Description:

The Performing Party will conduct water quality monitoring and provide details in the quarterly progress reports as prescribed in the FY2026-2027 CRP Guidance.

The Performing Party will complete the following subtasks:

Monitoring Description - The monitoring party will monitor a minimum of 93 routine stations per year. The sampling frequency and the types of parameter groups that are currently planned for collection in FY 2026 include: at a minimum, 19 monthly stations for field, select conventional parameters, and bacteria; 66 quarterly stations for field, select conventional parameters, and bacteria; five quarterly stations for field and bacteria only; seven bi-annual stations for field, select conventional and bacteria; and one station for field, 24-hr dissolved oxygen, bacteria and flow will be attempted if environmental conditions allow. The above monitoring will be routine monitoring with the objectives of collecting surface water data needed for conducting water quality assessments and identifying water quality trends. Two biological assessments will be attempted at the standard frequency of 2 if environmental conditions allow. Biological monitoring will be performed with the objective of providing baseline data on environmental conditions and to determine if criteria for ALU and dissolved oxygen criteria are being attained.

The Performing Party may also monitor approximately 11 additional stations in-kind as there are monitoring needs in excess of available funding. If additional funding is made available during the Contract period or if the allotted budget is underutilized, the in-kind stations may then be included in the Contract and reimbursement will be allowed. The sampling frequency and the types of parameter groups that are currently planned for collection in FY 2026 for in-kind stations include: 10 monthly stations for field, select conventional parameters, bacteria and flow; 1 station for field, select conventional parameters and bacteria.

For FY2027, the Performing Party will monitor at a similar level of effort as in FY2026. The actual number of sites, location, frequency, and parameters collected for FY 2027 will be included in the Performing Party's QAPP Appendix B update.

All monitoring will be completed according to the Performing Party QAPP, the *TCEQ Surface Water Quality Monitoring Procedures, Volume 1: Physical and Chemical Monitoring Methods* (RG-415) and the *TCEQ Surface Water Quality Monitoring Procedures, Volume 2: Methods for Collecting and Analyzing Biological Assemblage and Habitat Data* (RG-416).

Coordinated Monitoring Meeting - The Performing Party will hold an annual coordinated monitoring meeting as described in the FY2026-2027 CRP Guidance. Qualified monitoring organizations will be invited to attend the working meeting in which monitoring needs and purposes will be discussed segment by segment and station by station. Information from participants and stakeholders will be used to select stations and parameters that will enhance overall water quality monitoring coverage, eliminate duplication of effort, and address basin priorities. A summary of the changes to the monitoring schedule will be provided to the participants within two weeks of the meeting. Changes to the monitoring schedule will be entered into the statewide Coordinated Monitoring Schedule (<http://cms.lcra.org>) and communicated to meeting attendees. Changes to monitoring schedules that occur during the year will be entered into the Coordinated Monitoring Schedule and communicated to meeting attendees. All requirements related to meetings will be followed and required meetings will be conducted in-person or via TCEQ approved virtual format.

Monitoring Activities - Each progress report will include a description of activities including all types of monitoring performed, number of sampling events, and the types of monitoring conducted in the quarter. The Performing Party will complete and submit a monitoring activities report as an attachment to the progress report.

Deliverables and Dues Dates:

September 1, 2025 through August 31, 2026

- A. Conduct water quality monitoring, submit monitoring activities report, summarize activities, and submit with progress report—December 15, 2025; March 15 and June 15, 2026
- B. Coordinated Monitoring Meeting—between March 15 and April 30, 2026
- C. Coordinated Monitoring Meeting Summary of Changes—within 2 weeks following the meeting
- D. Email notification that Coordinated Monitoring Schedule updates are complete—May 31, 2026

September 1, 2026 through August 31, 2027

- A. Conduct water quality monitoring, submit monitoring activities report, summarize activities, and submit with progress report—September 15 and December 15, 2026; March 15 and June 15 and August 15, 2027
- B. Coordinated Monitoring Meeting—between March 15 and April 30, 2027
- C. Coordinated Monitoring Meeting Summary of Changes—within 2 weeks following the meeting
- D. Email notification that Coordinated Monitoring Schedule updates are complete—May 31, 2027

Sample Design Rationale FY 2026

The sample design is based on the legislative intent of CRP. Under the legislation, the Basin Planning Agencies have been tasked with providing data to characterize water quality conditions in support of the Texas Integrated Report of Surface Water Quality, and to identify significant long-term water quality trends. Based on Steering Committee input, achievable water quality objectives and priorities and the identification of water quality issues are used to develop work plans which are in accord with available resources. As part of the Steering Committee process, the Brazos River Authority coordinates closely with the TCEQ and other participants to ensure a comprehensive water monitoring strategy within the watershed. The most common water quality issues historically and currently in the basin, as with most of the State, are elevated bacteria and depressed dissolved oxygen. Bacterial and oxygen related water quality issues typically occur in small, rural streams with low to intermittent flow in the basin. In the upper region of the basin where natural brine springs contribute salts to the area, dissolved solids can be a water quality issue. Continued monitoring, as outlined in the current QAPP, can help elucidate the spatial and long-term trends in these parameters and others in the basin. Elucidating long-term trends is valuable for BRA and TCEQ decision making.

Changes made to the CRP monitoring schedule from, FY2025 to FY2026:

- 1202S – Bullhead Bayou/Steep Bank Creek – BRA will add for bacteria and field data collection at 17371 as TCEQ R12 is dropping.
- 1202T – Unnamed Tributary of Bullhead Bayou – BRA will add for bacteria and field data collection at 17382 as TCEQ R12 is dropping.
- 1202U – Alcorn Bayou– BRA will add for bacteria and field data collection at 17381 as TCEQ R12 is dropping.
- 1216A – Trimmier Creek – BRA will drop ALM data collection at 18754 as all needed data will have been collected by the end of FY25.
- 1216C – Pleasant Branch – BRA will drop bacteria data collection at 21689 as the City of Harker Heights

will take over this monitoring for their MS4 permit.

- 1216D – Unnamed Tributary of Trimmier Creek – BRA will drop bacteria data collection at 21690 as the City of Harker Heights will take over this monitoring for their MS4 permit.
- 1217A – Rocky Creek – BRA will add for ALM data collection at 11724 to provide baseline data on environmental conditions and to determine if an ALU is being attained.
- 1226 – North Bosque River – BRA will add for ALM data collection at 11951 as monitoring was not completed in FY25 to provide baseline data on environmental conditions and to determine if an ALU is being attained.
- 1242 – Brazos River – BRA will drop ALM data collection at 20833 as all needed data will have been collected by the end of FY25.
- 1242E – Little Brazos River – BRA will add for ALM data collection at 11591 to provide baseline data on environmental conditions and to determine if an ALU is being attained.
- 1243 – Salado Creek – BRA will drop ALM data collection at 20306 as all needed data will have been collected by the end of FY25.
- 1244 – Brushy Creek – BRA will drop ALM data collection at 12060 and 17374 as all needed data will have been collected by the end of FY25.
- 1248C – Mankins Branch – BRA will drop ALM data collection at 13497 as all needed data will have been collected by the end of FY25.
- 1250 – South Fork San Gabriel River –
 - BRA will drop ALM data collection at 20309 as all needed data will have been collected by the end of FY25.
 - BRA will add for ALM data collection at 12116 to provide baseline data on environmental conditions and to determine if an ALU is being attained.

Site Selection Criteria

This data collection effort involves monitoring routine water quality using procedures that are consistent with the TCEQ SWQM program. Some general guidelines are followed when selecting sampling sites, as outlined below, and discussed thoroughly in SWQM Procedures, Volumes I and II. Overall consideration is given to accessibility and safety. All monitoring activities have been developed in coordination with the CRP Steering Committee and with the TCEQ. The site selection criteria specified are those the TCEQ would like considered to produce data which is complementary to that collected by the state and which may be used in assessments, etc.

1. Locate stream sites so that samples can be safely collected from the centroid of flow. Centroid is defined as the midpoint of that portion of stream width which contains 50 percent of the total flow. If multiple potential sites on a stream segment are appropriate for monitoring, choose one that would best represent the water body, and not a site that displays unusual conditions or contaminant source(s). Avoid backwater areas or eddies when selecting a stream site.
2. At a minimum for reservoirs, locate sites near the dam (reservoirs) and in the major arms. Larger reservoirs might also include stations in the middle and upper (riverine) areas. Select sites that best represent the water body by avoiding coves and back water areas. A single monitoring site is considered representative of 25 percent of the total reservoir acres, but not more than 5,120 acres.
3. Monitoring sites are selected to maximize stream coverage or basin coverage. Very long segments may require more stations. As a rule of thumb, stream segments between 25 and 50 miles long require two stations, and longer than 50 miles require three or more depending on the existence of areas with significantly different sources of contamination or potential water quality concerns. Major hydrological features, such as the confluence of a major tributary or an instream dam, may also limit the spatial extent of an assessment based on one station.
4. Because historical water quality data can be very useful in assessing use attainment or impairment, it may be best to use sites that are on current or past monitoring schedules.
5. All classified segments (including reservoirs) should have at least one Monitoring site that adequately characterizes the water body, and monitoring should be coordinated with the TCEQ or other qualified monitoring entities reporting routine data to TCEQ.
6. Monitoring sites may be selected to bracket sources of pollution, influence of tributaries, changes in land uses, and hydrological modifications.

7. Sites should be accessible. When possible, stream sites should have a USGS or IBWC stream flow gauge. If not, it should be possible to conduct flow measurement during routine visits.

Monitoring Sites for FY 2026

Table B1.1 Sample Design and Schedule, FY 2026

A shaded comment field indicates the station and associated sampling are included in the budget for Contract 582-25-00054. Unshaded comment fields indicate that the station and associated sampling may be performed in-kind. If additional budget funds are made available during the Contract period or if the allotted budget is underutilized, the unshaded stations may then be included in the budget for Contract 582-26-00054.

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
BRAZOS RIVER 70 METERS DOWNSTREAM OF US 90A IN RICHMOND	11846	1202	12	12	BR	BR	RT	12	12	12	12					
BRAZOS RIVER AT FM 1458 NEAR SAN FELIPE	21816	1202	12	12	BR	BR	RT	12	12	12	12					
BRAZOS RIVER AT FM 1462 EAST BANK 4 MILES EAST OF WOODROW AND 7.4 MILES WEST OF ROSHARON	16355	1202	12	12	BR	BR	RT	12	12	12	12					
BRAZOS RIVER AT US 290 6.5 MILES NORTHWEST OF HEMPSTEAD	11850	1202	12	12	BR	BR	RT	12	12	12	12					
ALLENS CREEK AT FM 1458 NORTH OF WALLIS	11577	1202H	12	12	BR	BR	RT	4	4	4						
ALLENS CREEK AT MIXVILLE RD SOUTH OF SEALY	21753	1202H	12	12	BR	BR	RT	4	4	4						
BESSIES CREEK AT FM 1093 EAST OF FULBROOK ROAD AND SW OF FULSHEAR	21814	1202I	12	12	BR	BR	RT	4	4	4						
BULLHEAD BAYOU/STEEP BANK CREEK 156 METERS DOWNSTREAM OF LEXINGTON AVENUE IN SUGAR LAND	17372	1202S	12	12	BR	BR	RT	4		4						
BULLHEAD BAYOU/STEEP BANK CREEK AT WILLIAMS TRACE BOULEVARD/ SWEETWATER BOULEVARD IN SUGAR LAND	17371	1202S	12	12	BR	BR	RT	4		4						

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
UNNAMED TRIBUTARY OF BULLHEAD BAYOU AT AUSTIN PARKWAY IN SUGARLAND	17382	1202T	12	12	BR	BR	RT	4		4						
ALCORN BAYOU AT SWEETWATER BOULEVARD IN SUGARLAND	17381	1202U	12	12	BR	BR	RT	4		4						
BRAZOS RIVER 20 M OFF NORTH BANK AT FM 200 NORTHEAST OF GLEN ROSE	20213	1204	12	4	BR	BR	RT	12	12	12	12					
LAKE GRANBURY AT FM 51 NORTH OF GRANBURY 265 METERS WEST AND 69 METERS NORTH OF INTERSECTION OF FM 51 AND SIESTA COURT	11862	1205	12	4	BR	BR	RT	12	12	12						
LAKE GRANBURY IMMEDIATELY UPSTREAM OF ATCHISON TOPEKA AND SANTA FE RAILROAD 110 METERS UPSTREAM OF US377/EAST PEARL STREET EAST OF GRANBURY	20307	1205	12	4	BR	BR	RT	12	12	12						
LAKE GRANBURY NEAR DAM 102 METERS WEST AND 56 METERS NORTH OF NORTHERN EDGE OF DAM SITE AC USGS 322227097412101	11860	1205	12	4	BR	BR	RT	12	12	12						
BRAZOS RIVER AT FM 4 NORTH OF PALO PINTO	11864	1206	12	4	BR	BR	RT	12	12	12	12					
BRAZOS RIVER AT US 281 SOUTH OF MINERAL WELLS	11863	1206	12	4	BR	BR	RT	2	2	2						In support of the 79th TX Legislature's SB 1354
BRAZOS RIVER IMMEDIATELY DOWNSTREAM OF SOUTH SH 16	18748	1206	12	4	BR	BR	RT	2	2	2	2					In support of the 79th TX Legislature's SB 1354
BRAZOS RIVER IMMEDIATELY UPSTREAM FM 1189 SOUTH OF DENNIS	13543	1206	12	4	BR	BR	RT	12	12	12	12					

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
BRAZOS RIVER SOUTH BANK 1.74 KM DOWNSTREAM OF US 281 IN PALO PINTO COUNTY	18745	1206	12	4	BR	BR	RT	2	2	2						In support of the 79th TX Legislature's SB 1354
PALO PINTO CREEK IMMEDIATELY DOWNSTREAM OF FM 129 SOUTH OF BRAZOS	11074	1206D	12	4	BR	BR	RT	2	2	2						In support of the 79th TX Legislature's SB 1354
POSSUM KINGDOM RESERVOIR DEEP ELM CREEK ARM 597 METERS NORTH AND 880 METERS WEST OF INTERSECTION OF ANTHONY LOOP AND LEFTYS COURT	11868	1207	12	4	BR	BR	RT	12	12	12						
POSSUM KINGDOM RESERVOIR NEAR DAM 696 METERS WEST AND 221 METERS SOUTH OF NORTHERN EDGE OF DAM	11865	1207	12	4	BR	BR	RT	12	12	12						
POSSUM KINGDOM RESERVOIR NEAR END OF FM 2951 67 METERS NORTH AND 864 METERS WEST OF INTERSECTION OF FM 2951 AND SANBAR ROAD	11867	1207	12	4	BR	BR	RT	12	12	12						
POSSUM KINGDOM RESERVOIR NEAR JOHNSON BEND 437 METERS NORTH AND 429 METERS WEST OF INTERSECTION OF HELLS GATE LOOP AND HELLS POINT RD	11866	1207	12	4	BR	BR	RT	12	12	12						
BRAZOS RIVER 72 METERS DOWNSTREAM OF SH 67 2.0 MILES NE OF SOUTH BEND 2.81 KM DOWNSTREAM FROM THE CONFLUENCE WITH CLEAR FORK BRAZOS R	13641	1208	12	3	BR	BR	RT	12	12	12	12					
BRAZOS RIVER AT US 183/US 277 AT SEYMOUR	11871	1208	12	3	BR	BR	RT	12	12	12	12					

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
NAVASOTA RIVER AT GRIMES CR 162 5 MILES WEST OF FM 244 BETWEEN IOLA AND CARLOS	16398	1209	12	9	BR	BR	RT	4	4	4						
NAVASOTA RIVER IMMEDIATELY DOWNSTREAM OF SH 30 EAST OF COLLEGE STATION	11875	1209	12	9	BR	BR	RT	4	4	4						
NAVASOTA RIVER IMMEDIATELY DOWNSTREAM OF SH 6 NORTH OF NAVASOTA	11873	1209	12	9	BR	BR	RT	4	4	4						
NAVASOTA RIVER IMMEDIATELY DOWNSTREAM OF US 79 BETWEEN EASTERLY AND MARQUEZ	11877	1209	12	9	BR	BR	RT	4	4	4	4					
GIBBONS CREEK EAST 25 M UPSTREAM OF FM 244	18800	1209I	12	9	BR	BR	RT	4	4	4						
GIBBONS CREEK IMMEDIATELY DOWNSTREAM OF GRIMES CR 190 4.1 KM UPSTREAM OF THE CONFLUENCE WITH THE NAVASOTA RIVER	11756	1209I	12	9	BR	BR	RT	5		5	5	5				
NAVASOTA RIVER AT US 84 3.5 MILES UPSTREAM OF LAKE MEXIA	16391	1210A	12	9	BR	BR	RT	4	4	4						
YEGUA CREEK 377 METERS DOWNSTREAM OF FM 50 SOUTH OF CLAY	11880	1211	12	9	BR	BR	RT	4	4	4						
LITTLE RIVER AT MILAM CR 227/E 21ST ST NORTHEAST OF CAMERON	22084	1213	12	9	BR	BR	RT	12	12	12	12					
LITTLE RIVER IMMEDIATELY DOWNSTREAM OF SH 95 NEAR LITTLE RIVER ACADEMY	13546	1213	12	9	BR	BR	RT	4	4	4	4					

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
STILLHOUSE HOLLOW LAKE MID-LAKE AT LAMPASAS RIVER ARM APPROX 60 METERS UPSTREAM OF STILLHOUSE HOLLOW ROAD/FM 3481	11895	1216	12	9	BR	BR	RT	4	4	4						
STILLHOUSE HOLLOW LAKE NEAR DAM 441 METERS SOUTH AND 302 METERS WEST OF NORTHERN EDGE OF DAM SITE AC USGS 310129097315901	11894	1216	12	9	BR	BR	RT	4	4	4						
TRIMMIER CREEK IMMEDIATELY UPSTREAM OF CHAPARRAL ROAD WEST OF FM 3481	18754	1216A	12	9	BR	BR	RT	4	4	4						
ROCKY CREEK AT FM 963 AND APPROXIMATELY 1.26 KM UPSTREAM OF LAMPASAS RIVER NEAR OAKALLA	11724	1217A	12	11	BR	BR	BS	2	2		2	2	2	2	2	
ROCKY CREEK AT FM 963 AND APPROXIMATELY 1.26 KM UPSTREAM OF LAMPASAS RIVER NEAR OAKALLA	11724	1217A	12	11	BR	BR	RT	4	4	4						
NOLAN CREEK IMMEDIATELY UPSTREAM OF US 190 EAST OF NOLANVILLE	11907	1218	12	9	BR	BR	RT	4	4	4						
LITTLE NOLAN CREEK IMMEDIATELY DOWNSTREAM OF US190 BUSINESS AND 2.06 KILOMETERS UPSTREAM OF THE CONFLUENCE WITH SOUTH NOLAN CREEK IN KILLEEN TEXAS	21437	1218C	12	9	BR	BR	RT	4	4	4						
BELTON LAKE 629M NORTH AND 157M EAST OF THE BOAT RAMP AT WESTCLIFF PARK	20835	1220	12	9	BR	BR	RT	4	4	4						
BELTON LAKE IN OWL CREEK ARM 313 M NORTH AND 265 M WEST OF BOAT RAMP AT OWL CREEK PARK	18798	1220	12	9	BR	BR	RT	4	4	4						

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
BELTON RESERVOIR COWHOUSE CREEK ARM 88 METERS NORTH AND 954 METERS EAST OF THE INTERSECTION OF NOLAN CREEK ROAD AND LIBERTY HILL ROAD	11922	1220	12	9	BR	BR	RT	4	4	4						
BELTON RESERVOIR LEON RIVER ARM NEAR HEADWATERS 626 METERS N AND 288 METERS W OF INTERSECTION OF KUIKENDALL RD AND MC GREGOR PARK RD	11923	1220	12	9	BR	BR	RT	4	4	4						
BELTON RESERVOIR NEAR DAM 81 METERS NORTH AND 17 METERS WEST OF SOUTHERN EDGE OF DAM	11921	1220	12	9	BR	BR	RT	4	4	4						
COWHOUSE CREEK 71 METERS DOWNSTREAM OF FM 116 SOUTHWEST OF GATESVILLE	11805	1220A	12	9	BR	BR	RT	4	4	4	4					
LEON RIVER 18 METERS UPSTREAM OF CORYELL CR 183 NORTHEAST OF LEVITA	11929	1221	12	9	BR	BR	RT	4	4	4						
LEON RIVER AT HAMILTON COUNTY ROAD 109	18781	1221	12	3	BR	BR	RT	4	4	4						
LEON RIVER AT HAMILTON CR 203 NORTH OF HAMILTON	20905	1221	12	9	BR	BR	RT	12	12	12						
LEON RIVER AT HAMILTON CR 431 1.6 KM DOWNSTREAM OF SH 36 SOUTHWEST OF JONESBORO	11930	1221	12	9	BR	BR	RT	12	12	12						
LEON RIVER IMMEDIATELY DOWNSTREAM OF US 67/ US 377 DOWNSTREAM LAKE PROCTOR	11934	1221	12	3	BR	BR	RT	12	12	12	12					

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
RESLEY CREEK AT COMANCE CR 394 740 METERS UPSTREAM OF THE CONFLUENCE WITH THE LEON RIVER	11808	1221A	12	3	BR	BR	RT	4	4	4						
RESLEY CREEK AT FM 2823 WEST OF CARLTON C704	17377	1221A	12	3	BR	BR	RT	4	4	4						
SOUTH LEON RIVER 300 METERS DOWNSTREAM OF COMANCHE CR 260 EAST OF GUSTINE	20390	1221B	12	3	BR	BR	RT	4	4	4						
PECAN CREEK AT SH 22 EAST OF HAMILTON	17547	1221C	12	9	BR	BR	RT	4	4	4						
PLUM CREEK 10 M DOWNSTREAM OF CORYELL CR 106 NEAR LEVITA	18405	1221E	12	9	BR	BR	RT	4	4	4						
CORYELL CREEK 51 METERS DOWNSTREAM OF FM 107 1.9 KM UPSTREAM OF THE CONFLUENCE WITH THE LEON RIVER	11804	1221G	12	9	BR	BR	RT	4	4	4						
PROCTOR LAKE COPPERAS CREEK ARM 460 METERS NORTH AND 2.04 KILOMETERS EAST OF INTERSECTION OF COMANCE CR 410A AND COMANCHE CR 407	11937	1222	12	3	BR	BR	RT	2	2	2						
PROCTOR LAKE IN LEON AND SABANA RIVER ARM 2.43 KM NORTH AND 1.23 KM EAST OF INTERSECTION OF COMANCHE CR 424 AND FM 2318	11936	1222	12	3	BR	BR	RT	2	2	2						
PROCTOR LAKE NEAR DAM FLOODGATE 911 METERS NORTH AND 940 METERS EAST OF INTERSECTION OF FM 2861 AND COMANCHE CR 418C	11935	1222	12	3	BR	BR	RT	2	2	2						

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
NORTH BOSQUE RIVER AT COOPERS CROSSING ROAD WEST OF CHINA SPRING	11951	1226	12	9	BR	BR	BS	2	2		2	2	2	2	2	
NORTH BOSQUE RIVER AT COOPERS CROSSING ROAD WEST OF CHINA SPRING	11951	1226	12	9	BR	BR	RT	4	4	4						
MERIDIAN CREEK AT SH 6 2.5 MILES NORTHWEST OF CLIFTON	14908	1226C	12	9	BR	BR	RT	4	4	4						
NOLAN RIVER 75 METERS UPSTREAM OF FM 933 IN BLUM	11967	1227	12	9	BR	BR	RT	4	4	4	4					
NOLAN RIVER IMMEDIATELY UPSTREAM OF FM 916 WEST OF RIO VISTA	11971	1227	12	4	BR	BR	RT	4	4	4						
PALUXY RIVER LOW WATER CROSSING OFF OF VAN ZANDT ROAD NEAR SH 144 IN GLEN ROSE	20232	1229	12	4	BR	BR	RT	4	4	4						
BRAZOS RIVER AT SH 105 WEST OF NAVASOTA	12030	1242	12	9	BR	BR	RT	12	12	12						
BRAZOS RIVER AT SH 21 11 MILES NORTHEAST OF CALDWELL	15767	1242	12	9	BR	BR	RT	12	12	12	12					
BRAZOS RIVER IMMEDIATELY DOWNSTREAM OF FM 413 NORTHEAST OF ROSEBUD	12032	1242	12	9	BR	BR	RT	12	12	12	12					
BRAZOS RIVER IMMEDIATELY UPSTREAM OF SH 6 SOUTHEAST OF WACO	12038	1242	12	9	BR	BR	RT	12	12	12	12					
COTTONWOOD BRANCH AT THE CONFLUENCE WITH STILL CREEK 50 METERS DOWNSTREAM OF SH 21	17598	1242B	12	9	BR	BR	RT	4	4	4						
STILL CREEK AT FM 2818 WEST OF BRYAN	17378	1242C	12	9	BR	BR	RT	4	4	4						
STILL CREEK AT SH 21 WEST OF BRYAN	16882	1242C	12	9	BR	BR	RT	4	4	4						

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
THOMPSONS CREEK IMMEDIATELY UPSTREAM OF SILVERHILL ROAD 765 METERS UPSTREAM OF SH 47 WEST OF BRYAN	16396	1242D	12	9	BR	BR	RT	4	4	4						
LITTLE BRAZOS RIVER IMMEDIATELY UPSTREAM OF SH 21 WEST OF BRYAN	11591	1242E	12	9	BR	BR	BS	2	2		2	2	2	2	2	
LITTLE BRAZOS RIVER IMMEDIATELY UPSTREAM OF SH 21 WEST OF BRYAN	11591	1242E	12	9	BR	BR	RT	4	4	4	4					
POND CREEK AT FM 2027 4.0 KILOMETERS SOUTH OF BAILEYVILLE	16406	1242F	12	9	BR	BR	RT	4	4	4						
CAMPBELLS CREEK AT SH 6/US 190 14 MILES SOUTH OF HEARNE	16395	1242I	12	9	BR	BR	RT	4		4						
DEER CREEK IMMEDIATELY DOWNSTREAM OF SH 320 WEST OF MARLIN	11723	1242J	12	9	BR	BR	RT	4	4	4						
MUD CREEK AT SH 6 4.5 MI NW OF HEARNE	16402	1242K	12	9	BR	BR	RT	4		4						
PIN OAK CREEK AT SH 6/US 190 2 MILES SOUTHEAST OF HEARNE	16401	1242L	12	9	BR	BR	RT	4		4						
SPRING CREEK AT SH 6/US 190 8 MILES SOUTH OF HEARNE	16394	1242M	12	9	BR	BR	RT	4		4						
BIG CREEK IMMEDIATELY UPSTREAM OF SH 6 SOUTH OF MARLIN	16400	1242P	12	9	BR	BR	RT	4		4						
SALADO CREEK 75 METERS DOWNSTREAM OF FM 2268 IN SALADO	12051	1243	12	9	BR	BR	RT	4	4	4						
BRUSHY CREEK IMMEDIATELY DOWNSTREAM OF CHISHOLM TRAIL ROAD	12068	1244	12	11	BR	BR	RT	4	4	4						

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
BRUSHY CREEK IMMEDIATELY DOWNSTREAM OF FM 685	12060	1244	12	11	BR	BR	RT	4	4	4						
BRUSHY CREEK IMMEDIATELY UPSTREAM OF FM 973 APPROXIMATELY 7.35 KM NORTHWEST OF COUPLAND, TX	22395	1244	12	11	BR	BR	RT	4	4	4	4					
WASP CREEK AT SH 317 APPROXIMATELY 0.8 KM SOUTH OF CRAWFORD	18802	1246E	12	9	BR	BR	RT	4	4	4						
GRANGER LAKE IN SAN GABRIEL RIVER ARM NEAR HEADWATERS 7.22 KILOMETERS DOWNSTREAM OF SH 95	12096	1247	12	11	BR	BR	RT	4	4	4						
GRANGER LAKE IN WILLIS CREEK ARM 960 METERS NORTH AND 1.91 KM EAST OF INTERSECTION OF WILLIAMSON CR 348 AND CR 389	12097	1247	12	11	BR	BR	RT	4	4	4						
GRANGER LAKE NEAR DAM 1.44 KILOMETERS NORTH AND 190 METERS WEST OF SOUTHERN EDGE OF DAM	12095	1247	12	11	BR	BR	RT	4	4	4						
WILLIS CREEK AT WILLIAMSON CR 236 WEST OF GRANGER 635 METERS EAST OF THE INTERSECTION OF WILLIAMSON CR 335 AND WILLIAMSON CR 326	20305	1247A	12	11	BR	BR	RT	4	4	4						
SAN GABRIEL/NORTH FORK SAN GABRIEL RIVER AT WILLIAMSON CR 366 4.84 KILOMETERS UPSTREAM OF SH 95	12099	1248	12	11	BR	BR	RT	4	4	4						
SAN GABRIEL/NORTH FORK SAN GABRIEL RIVER IMMEDIATELY DOWNSTREAM OF SH 29 EAST OF GEORGETOWN	12102	1248	12	11	BR	BR	RT	4	4	4						

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
SAN GABRIEL/NORTH FORK SAN GABRIEL RIVER NORTH FORK IMMEDIATELY DOWNSTREAM OF IH 35 IN GEORGETOWN	12108	1248	12	11	BR	BR	RT	4	4	4						
BERRY CREEK IMMEDIATELY DOWNSTREAM OF FM 971 2 MILES EAST OF IH 35	13496	1248A	12	11	BR	BR	RT	4	4	4						
MANKINS BRANCH AT WILLIAMSON CR 100 IMMEDIATELY UPSTREAM OF THE CONFLUENCE WITH THE SAN GABRIEL RIVER	13497	1248C	12	11	BR	BR	RT	4	4	4						
LAKE GEORGETOWN NEAR DAM 68 METERS NORTH AND 88 METERS EAST OF SOUTHWEST EDGE OF DAM	12111	1249	12	11	BR	BR	RT	4	4	4						
LAKE GEORGETOWN NEAR HEADWATERS IN THE NORTH SAN GABRIEL ARM 305 METERS SOUTH AND 1.05 KILOMETERS WEST FROM THE INTERSECTION OF WILLIAMSON CR 262 AND PARK ROAD 8	12113	1249	12	11	BR	BR	RT	4	4	4						
SOUTH FORK SAN GABRIEL RIVER 1.44 KM NORTH AND 1.80 KM WEST OF THE INTERSECTION OF WEIR RANCH ROAD AND LEANDER RANCH ROAD / RR 2243 AT WEIR PIT ROCK QUARRY IN WILLIAMSON COUNTY	20309	1250	12	11	BR	BR	RT	4	4	4	4					
SOUTH FORK SAN GABRIEL RIVER AT RONALD REAGAN BLVD NE OF LEANDER	21739	1250	12	11	BR	BR	RT	4	4	4	4					
SOUTH FORK SAN GABRIEL RIVER AT US 183	12116	1250	12	11	BR	BR	RT	4	4	4	4					
SOUTH FORK SAN GABRIEL RIVER AT US 183	12116	1250	12	11	BR	BR	BS	2	2		2	2	2	2	2	

Site Description	Station ID	Waterbody ID	Basin	Region	SE	CE	MT	Field	Conv	Bacteria	Flow	24 hr DO	AqHab	Benthics	Nekton	Comments
LAKE LIMESTONE AT CONFLUENCE OF NAVASOTA RIVER AND BIG CREEK ARMS 1.33 KM S AND 1.39 KM EAST OF INTERSECTION OF LCR 752 AND 3D RCH RD	12125	1252	12	9	BR	BR	RT	12	12	12						
LAKE LIMESTONE AT FM 3371 696 METERS NORTH AND 430 METERS EAST OF INTERSECTION OF FM 3371 AND PARK 2 RD SITE DC USGS 312622096224201	13970	1252	12	9	BR	BR	RT	12	12	12						
LAKE LIMESTONE IN LAMBS CREEK ARM 2.19 KILOMETERS DOWNSTREAM OF FM 1512 NEAR LCR 893	12124	1252	12	9	BR	BR	RT	12	12	12						
LAKE LIMESTONE NEAR DAM 572 METERS NORTH AND 2.28 KILOMETERS EAST OF INTERSECTION OF WINDING WAY ROAD AND BRAZOS RIVER AUTHORITY ROAD	12123	1252	12	9	BR	BR	RT	12	12	12						
UPPER NAVASOTA RIVER 81 METERS DOWNSTREAM OF SH 164 EAST OF GROESBECK	12126	1253	12	9	BR	BR	RT	4	4	4						
AQUILLA CREEK IMMEDIATELY UPSTREAM OF FM 933 NORTHWEST OF WACO	11593	1256A	12	9	BR	BR	RT	4	4	4						
BRAZOS RIVER IMMEDIATELY UPSTREAM OF FM 2114 SOUTHEAST OF LAGUNA PARK	12044	1257	12	9	BR	BR	RT	12	12	12	12					
LEON RIVER IMMEDIATELY DOWNSTREAM OF FM 1829 SOUTHEAST OF NORTH FORT HOOD	11925	1259	12	9	BR	BR	RT	12	12	12						

Table B1.2 Specific parameters collected and frequency to accompany sample design and schedule, FY 2026.

SEGMENT	RUN/STATION ID	DESCRIPTION	PARAMETER															
			Chloride	Sulfate	Nitrite	Nitrate	OrthoP	TSS	TDS	Turbidity	Chl a	E. coli	Enteroc	TKN	TP	Ammonia	Flow	24-hr DO
	Upper Brazos																	
1208	11871	Brazos River at US 183 near Seymour	12	12	12	12	12	12	12	12	12	12		12	12	12		12
1208	13641	Brazos River at SH 67 NE of South Bend	12	12	12	12	12	12	12	12	12	12		12	12	12	12	12
1206	11864	Brazos River at FM 4 N of Palo Pinto	12	12	12	12	12	12			12	12	12		12	12	12	12
1206	13543	Brazos River at FM 1189 in Dennis	12	12	12	12	12	12	12	12	12	12		12	12	12	12	12
1204	20213	Brazos River at FM 200 NE of Glen Rose	12	12	12	12	12	12	12	12	12	12		12	12	12	12	12
1257	12044	Brazos River at FM 2114 SE of Laguna Park	12	12	12	12	12	12	12	12	12	12		12	12	12	12	12
	PK																	
1207	11868	Possum Kingdom Lake at Deep Elm arm	12	12	12	12	12	12			12		12					
1207	11867	Possum Kingdom Lake at FM 2951 (Sandy Beach)	12	12	12	12	12	12			12		12					
1207	11866	Possum Kingdom Lake at Johnson Road	12	12	12	12	12	12			12		12					
1207	11865	Possum Kingdom Lake at Morris Shepard Dam	12	12	12	12	12	12	12	12	12	12		12	12	12		
	Granbury																	
1205	11860	Lake Granbury at De Cordova Dam	12	12	12	12	12	12	12	12	12	12		12	12	12		
1205	11862	Lake Granbury at FM 51	12	12	12	12	12	12			12	12	12		12	12	12	
1205	20307	Lake Granbury at US Business 377	12	12	12	12	12	12			12	12	12		12	12	12	
	John Graves																	
1206	18748	Brazos River at SH 16 near Graford	2	2	2	2	2	2			2		2					2
1206D	11074	Palo Pinto Creek at FM 129 S of Brazos	2	2	2	2	2	2	2	2	2		2					
1206	11863	Brazos River at US 281 S of Mineral Wells	2	2	2	2	2	2	2	2	2		2					
1206	18745	Brazos River dwnstr. of US 281 at Rivercrest Campground	2	2	2	2	2	2			2		2					
	Lake Proctor																	
1222	11935	Lake Proctor at Dam	2	2	2	2	2	2	2	2	2	2		2	2	2		
1222	11936	Lake Proctor at Leon River arm	2	2	2	2	2	2			2		2					
1222	11937	Lake Proctor at Copperas Creek arm	2	2	2	2	2	2			2		2					
	Resley Creek																	

SEGMENT	RUN/STATION ID	DESCRIPTION	PARAMETER																
			Chloride	Sulfate	Nitrite	Nitrate	OrthoP	TSS	TDS	Turbidity	Chl a	E. coli	Entero	TKN	TP	Ammonia	Flow	24-hr DO	
1221G	11804	Coryell Creek at FM 107 E of Gatesville	4	4	4	4	4	4		4		4							
1221E	18405	Plum Creek at CR 106 W of Levita	4	4	4	4	4	4		4		4							
1221	11929	Leon River at CR 198 NE of Levita	4	4	4	4	4	4		4	-	4		-	-				
1221C	17547	Pecan Creek at SH 22 E of Hamilton	4	4	4	4	4	4		4		4							
1221B	20390	South Leon River near Comanche CR 260 E of Gustine	4	4	4	4	4	4		4		4							
1221	18781	Leon River at CR 109 NW of Hamilton	4	4	4	4	4	4		4		4							
1221A	11808	Resley Creek at CR 394 E of Gustine	4	4	4	4	4	4		4	4	4		4	4				
1221A	17377	Resley Creek at FM 2823 W of Carlton	4	4	4	4	4	4		4	4	4		4	4				
	Leon River																		
1221	11934	Leon River at US 67 / US 377 below Lake Proctor	12	12	12	12	12	12		12	12	12	-	12	12		12		
1221	20905	Leon River at CR 203 N of Hamilton	12	12	12	12	12	12		12	12	12		12	12				
1221	11930	Leon River at CR 431 SW of Jonesboro	12	12	12	12	12	12		12		12							
1259	11925	Leon River at FM 1829 SE of Gatesville	12	12	12	12	12	12	12	12	12	12		12	12	12			
	North Bosque																		
1256A	11593	Aquilla Creek at FM 933 SE of Gholson	4	4	4	4	4	4	4	4	4	4		4	4	4			
1227	11967	Nolan River at FM 933 N of Blum	4	4	4	4	4	4	4	4	4	4		4	4	4	4		
1227	11971	Nolan River at FM 916 W of Rio Vista	4	4	4	4	4	4		4		4							
1229	20232	Paluxy River at low water crossing off Van Zandt Rd. in Glen Rose	4	4	4	4	4	4		4		4							
1226C	14908	Meridian Creek at SH 6 NW of Clifton	4	4	4	4	4	4		4		4							
1226	11951	North Bosque River at Coopers Crossing W of China Spring	4	4	4	4	4	4	4	4	4	4		4	4	4			
1226	11951	North Bosque River at Coopers Crossing W of China Spring	2	2	2	2	2	2	2	2	2		2	2	2	2	2	2	
1246E	18802	Wasp Creek at SH 317 S of Crawford	4	4	4	4	4	4		4		4							
	Sulphur Creek																		
1220A	11805	Cowhouse Creek at FM 116 near Pidcoke	4	4	4	4	4	4		4		4					4		
1217A	11724	Rocky Creek at FM 963 near Oakalla	4	4	4	4	4	4		4		4							
1217A	11724	Rocky Creek at FM 963 near Oakalla	2	2	2	2	2	2	2	2	2		2	2	2	2	2	2	
1216A	18754	Trimmier Creek at Chaparral Rd. S of Harker Heights	4	4	4	4	4	4		4		4							
1218	11907	Nolan Creek (South Nolan Creek) at US 190 E of Nolanville	4	4	4	4	4	4		4		4							

SEGMENT	RUN/STATION ID	DESCRIPTION	PARAMETER															
			Chloride	Sulfate	Nitrite	Nitrate	OrthoP	TSS	TDS	Turbidity	Chl a	E. coli	Entero	TKN	TP	Ammonia	Flow	24-hr DO
1218C	21437	Little Nolan Creek at US Business 190 in Killeen	4	4	4	4	4	4		4		4						
	Stillhouse																	
1216	11894	Stillhouse Hollow Lake at Dam	4	4	4	4	4	4	4	4	4	4		4	4	4		
1216	11895	Stillhouse Hollow Lake at Dam	4	4	4	4	4	4		4	4	4		4	4			
	Belton																	
1220	11923	Belton Lake at Leon River arm	4	4	4	4	4	4		4		4						
1220	11922	Belton Lake at Cowhouse Creek arm	4	4	4	4	4	4		4		4						
1220	11921	Belton Lake at Dam	4	4	4	4	4	4	4	4	4		4	4	4			
1220	18798	Belton Lake at Owl Creek arm	4	4	4	4	4	4		4		4						
1220	20835	Belton Lake E of Westcliff	4	4	4	4	4	4		4		4						
	Georgetown/Granger																	
1247	12095	Granger Lake at Dam	4	4	4	4	4	4	4	4	4		4	4	4			
1247	12097	Granger Lake at Willis Creek arm	4	4	4	4	4	4		4	4	4		4	4			
1247	12096	Granger Lake at San Gabriel River arm	4	4	4	4	4	4		4	4	4		4	4			
1249	12113	Lake Georgetown at San Gabriel River arm	4	4	4	4	4	4		4	4	4		4	4			
1249	12111	Lake Georgetown at Dam	4	4	4	4	4	4	4	4	4		4	4	4			
	San Gabriel																	
1213	13546	Little River at SH 95 S of Little River Academy	4	4	4	4	4	4	4	4	4		4	4	4	4		
1247A	20305	Willis Creek at CR 326 W of Granger	4	4	4	4	4	4		4	4	4		4	4	4		
1248	12099	San Gabriel River at CR 366, 4.5 km upstr. of SH 95	4	4	4	4	4	4	4	4	4		4	4	4			
1248	12102	San Gabriel River at SH 29 E of Georgetown	4	4	4	4	4	4	4	4	4		4	4	4			
1248C	13497	Mankins Branch at CR 100 W of Jonah	4	4	4	4	4	4	4	4	4		4	4	4			
1248A	13496	Berry Creek at FM 971 W of Weir	4	4	4	4	4	4		4		4						
1243	12051	Salado Creek near FM 2268, below Serena Spring	4	4	4	4	4	4		4		4						
	SFSGR																	
1250	20309	South Fork San Gabriel River at Wier Pit W of Georgetown	4	4	4	4	4	4	4	4	4		4	4	4	4		
1250	12116	South Fork San Gabriel River at US 183 N of Leander	4	4	4	4	4	4		4	4	4		4	4	4	4	
1250	12116	South Fork San Gabriel River at US 183 N of Leander	2	2	2	2	2	2	2	2	2		2	2	2	2	2	2

SEGMENT	RUN/STATION ID	DESCRIPTION	PARAMETER															
			Chloride	Sulfate	Nitrite	Nitrate	OrthoP	TSS	TDS	Turbidity	Chl a	E. coli	Enteroc	TKN	TP	Ammonia	Flow	24-hr DO
1250	21739	South Fork San Gabriel River at Ronald Reagan Blvd. NE of Leander	4	4	4	4	4	4		4	4	4		4	4	4	4	
	Brushy Creek																	
1244	12068	Brushy Creek at Chisholm Trail Rd. in Round Rock	4	4	4	4	4	4	4	4	4	4		4	4	4		
1244	12060	Brushy Creek at FM 685 SW of Hutto	4	4	4	4	4	4	4	4	4	4		4	4	4		
1248	12108	North Fork San Gabriel River at Austin Ave. in Georgetown	4	4	4	4	4	4	4	4	4	4		4	4	4		
1244	22395	Brushy Creek at FM 973 near Coupland	4	4	4	4	4	4	4	4	4	4		4	4	4	4	
	Navasota																	
1210A	16391	Navasota River at US 84 above Lake Mexia	4	4	4	4	4	4		4	4	4		4	4	4		
1253	12126	Navasota River at SH 164 E of Groesbeck	4	4	4	4	4	4	4	4	4	4		4	4	4		
1209	11877	Navasota River at US 79 SW of Marquez	4	4	4	4	4	4	4	4	4	4		4	4	4	4	
1242F	16406	Pond Creek at FM 2027 S of Baileyville	4	4	4	4	4	4		4		4						
1242J	11723	Deer Creek at SH 320 W of Marlin	4	4	4	4	4	4		4		4						
	Limestone																	
1252	12123	Lake Limestone at Sterling C. Robertson Dam	12	12	12	12	12	12	12	12	12	12		12	12	12		
1252	12124	Lake Limestone at Lambs Creek arm	12	12	12	12	12	12		12	12	12		12	12	12		
1252	12125	Lake Limestone at Big Creek arm	12	12	12	12	12	12		12	12	12		12	12	12		
1252	13970	Lake Limestone at FM 3371	12	12	12	12	12	12		12	12	12		12	12	12		
	College Station East																	
1209	11873	Navasota River at SH 6 NW of Navasota	4	4	4	4	4	4	4	4	4	4		4	4	4		
1209I	18800	Gibbons Creek East at FM 244 SE of Carlos	4	4	4	4	4	4		4		4						
1209I	11756	Gibbons Creek at Grimes CR 190										5					5	5
1209	11875	Navasota River at SH 30 NE of College Station	4	4	4	4	4	4		4	4	4		4	4	4		
1209	16398	Navasota River at Grimes CR 162	4	4	4	4	4	4		4	4	4		4	4	4		
	College Station West																	
1242E	11591	Little Brazos River at SH 21 SW of Bryan	4	4	4	4	4	4		4	4	4		4	4	4	4	
1242E	11591	Little Brazos River at SH 21 SW of Bryan	2	2	2	2	2	2	2	2	2			2	2	2	2	2
1242D	16396	Thompsons Creek at Silver Hill Rd. SW of Bryan	4	4	4	4	4	4		4	4	4			4	4		
1242C	16882	Still Creek at SH 21 W of Bryan	4	4	4	4	4	4		4		4						

SEGMENT	RUN/STATION ID	DESCRIPTION	PARAMETER																
			Chloride	Sulfate	Nitrite	Nitrate	OrthoP	TSS	TDS	Turbidity	Chl a	E. coli	Enteroc	TKN	TP	Ammonia	Flow	24-hr DO	
1242B	17598	Cottonwood Branch near SH 21 W of Bryan	4	4	4	4	4	4		4		4							
1242C	17378	Still Creek at FM 2818 W of Bryan	4	4	4	4	4	4		4		4							
	HWY 6																		
1242I	16395	Cambells Creek at SH 6/ US190										4							
1242M	16394	Spring Creek at SH6/US190										4							
1242L	16401	Pin Oak Creek at SH 6/US 190										4							
1242K	16402	Mud Creek at SH 6										4							
1242P	16400	Big Creek at SH 6										4							
	Allens Creek																		
1202I	21814	Bessie's Creek at FM 1093 SW of Fulshear	4	4	4	4	4	4		4	4	4		4	4	4			
1202H	11577	Allens Creek at FM 1458 NE of Wallis	4	4	4	4	4	4		4	4	4		4	4	4			
1202H	21753	Allens Creek at Mixville Rd. S of Sealy	4	4	4	4	4	4		4	4	4		4	4	4			
1211	11880	Yegua Creek at FM 50 N of Independence	4	4	4	4	4	4	4	4	4	4		4	4	4			
1202S	17371	Bullhead Bayou/Steep Bank Creek at Williams Trace Blvd in Sugarland										4							
1202S	17372	Bullhead Bayou/Steep Bank Creek near Lexington Ave in Sugarland										4							
1202T	17382	Unnamed Trib of Bullhead Bayou at Austin Parkway in Sugarland										4							
1202U	17381	Alcorn Bayou at Sweetwater Blvd in Sugarland										4							
	Highbank																		
1242	15767	Brazos River at SH 21 SW of Bryan	12	12	12	12	12	12		12	12	12		12	12	12	12		
1213	22084	Little River at Milam CR 227 NE of Cameron	12	12	12	12	12	12	12	12	12	12		12	12	12	12		
1242	12032	Brazos River at FM 413 S of Highbank	12	12	12	12	12	12	12	12	12	12		12	12	12	12		
1242	12038	Brazos River at Loop 340 E of Waco	12	12	12	12	12	12		12	12	12		12	12	12	12		
	Richmond																		
1202	16355	Brazos River at FM 1462 W of Rosharon	12	12	12	12	12	12	12	12	12	12		12	12	12	12		
1202	21816	Brazos River at FM 1458 near San Felipe	12	12	12	12	12	12	12	12	12	12		12	12	12	12		
1202	11846	Brazos River at US 90A in Richmond	12	12	12	12	12	12	12	12	12	12		12	12	12	12		
1202	11850	Brazos River at US 290 NW of Hempstead	12	12	12	12	12	12	12	12	12	12		12	12	12	12		
1242	12030	Brazos River at SH 105 SW of Navasota	12	12	12	12	12	12		12	12	12		12	12	12			

SEGMENT	RUN/STATION ID	DESCRIPTION	PARAMETER															
			Chloride	Sulfate	Nitrite	Nitrate	OrthoP	TSS	TDS	Turbidity	Chl a	E. coli	Enteroc	TKN	TP	Ammonia	Flow	24-hr DO

	biological site
X	24-hr DO site

Appendix C: Station Location Maps

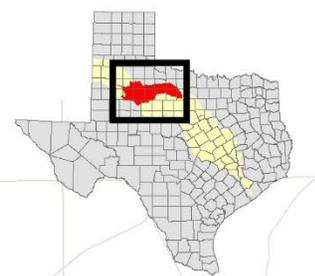
Station Location Maps

Maps of stations monitored by the Brazos River Authority are provided below. The maps were generated by the Brazos River Authority. This product is for informational purposes and may not have been prepared for or be suitable for legal, engineering, or surveying purposes. It does not represent an on-the-ground survey and represents only the approximate relative location of property boundaries. For more information concerning this map, contact Jenna Olson at (254) 761-3149.

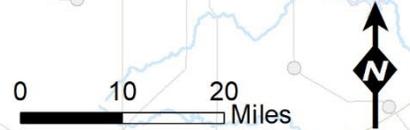
Watershed of the Salt and Double Mountain Forks of the Brazos River

FY26

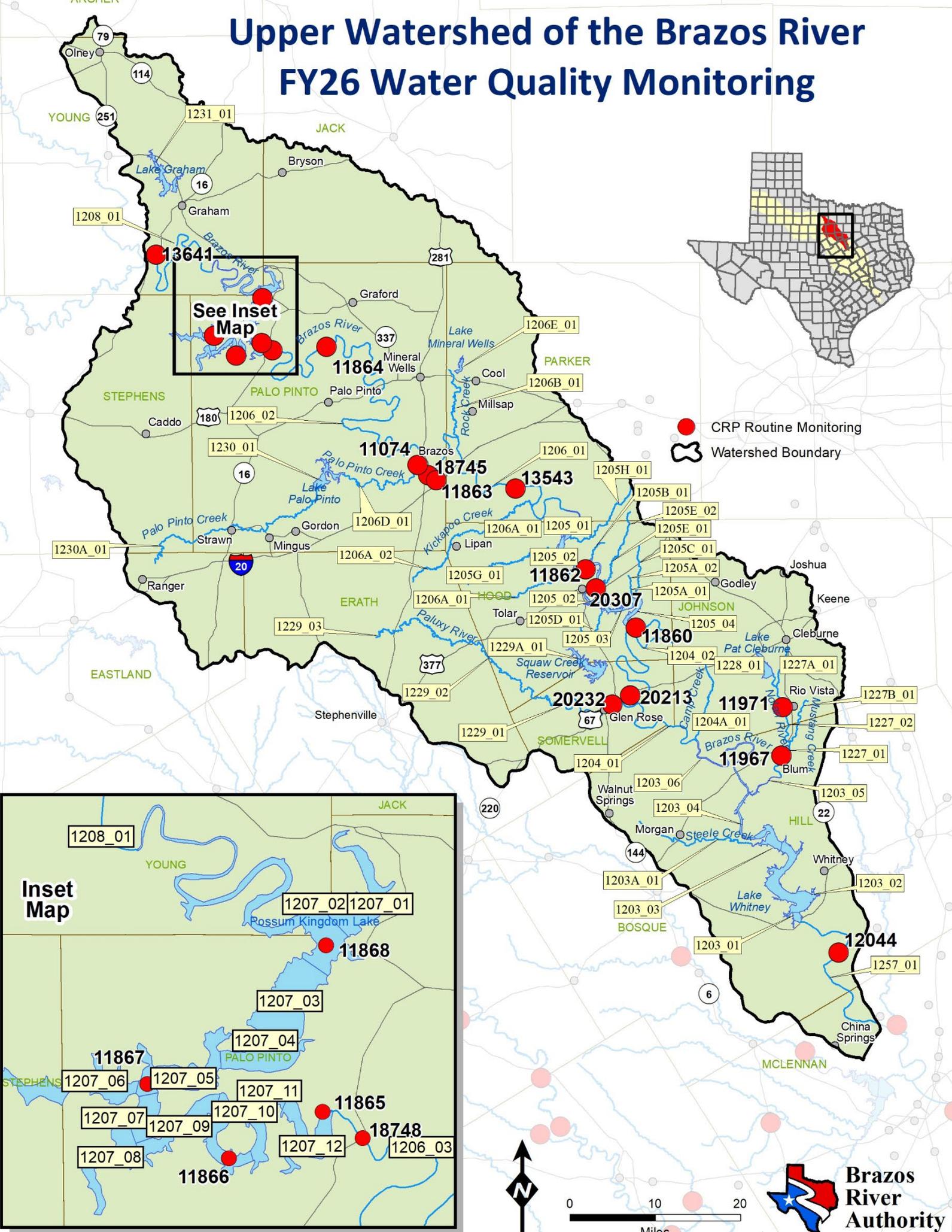
Water Quality Monitoring



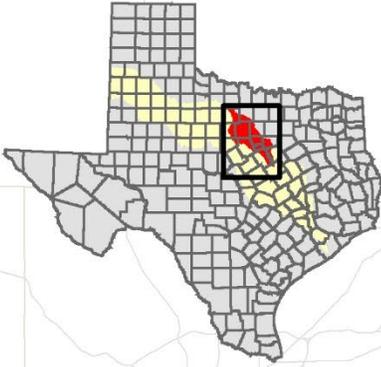
● CRP Routine Monitoring
⬭ Watershed Boundary



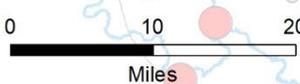
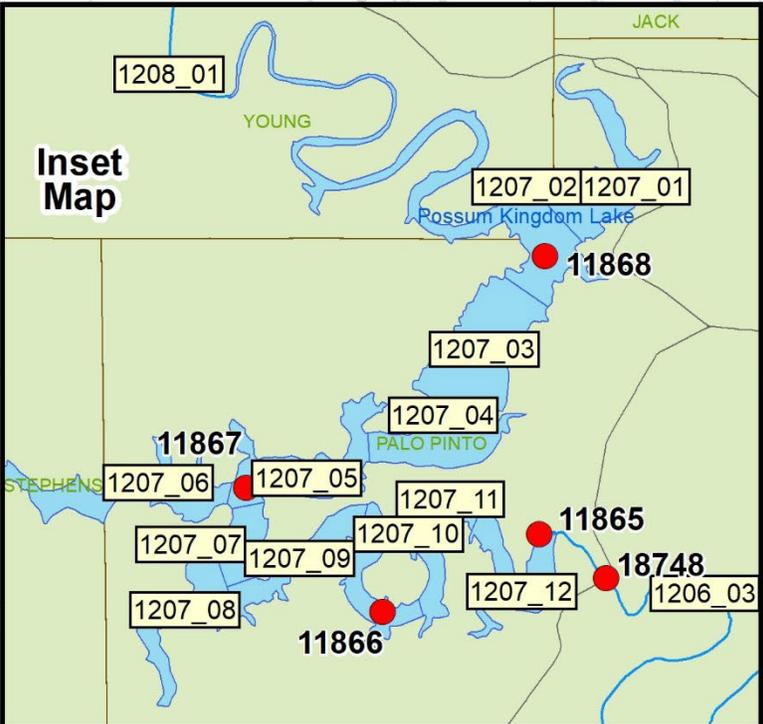
Upper Watershed of the Brazos River FY26 Water Quality Monitoring



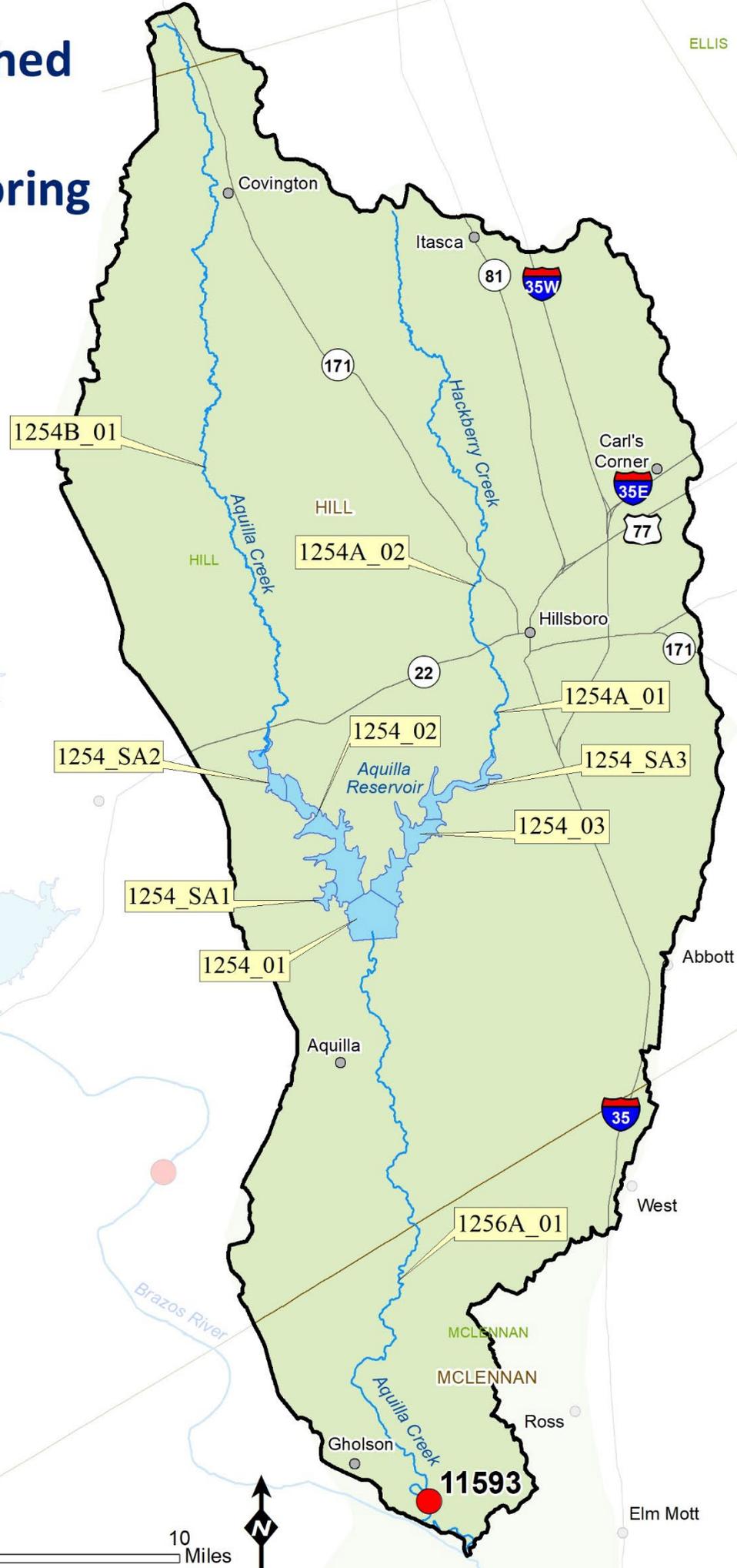
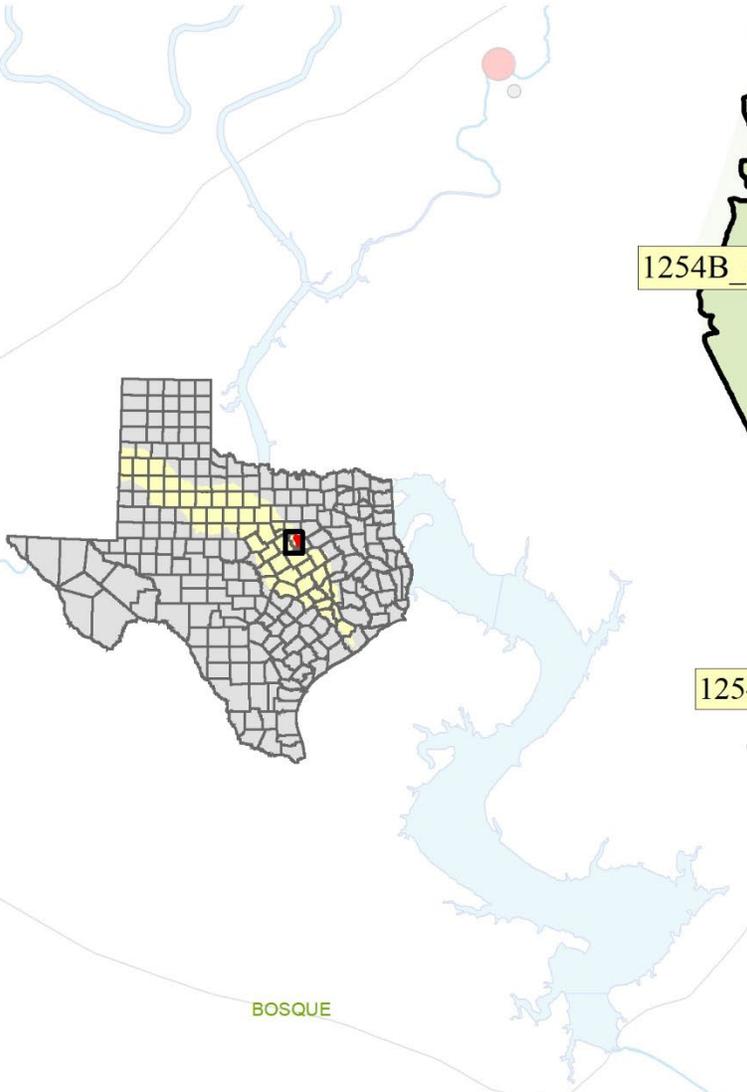
See Inset Map



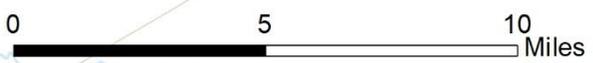
● CRP Routine Monitoring
 ⬭ Watershed Boundary



Aquilla Creek Watershed FY26 Water Quality Monitoring



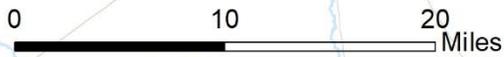
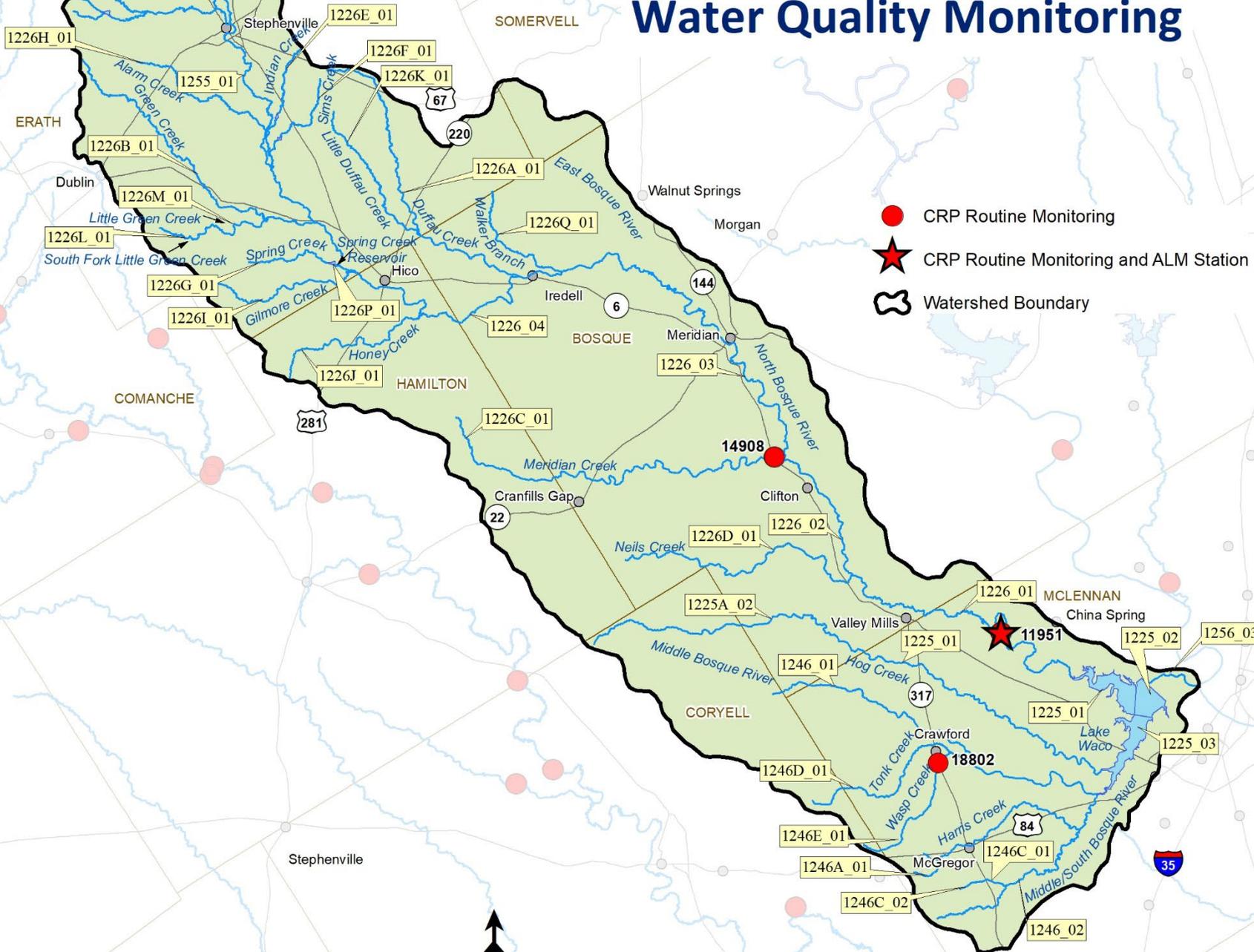
- CRP Routine Monitoring
- Watershed Boundary



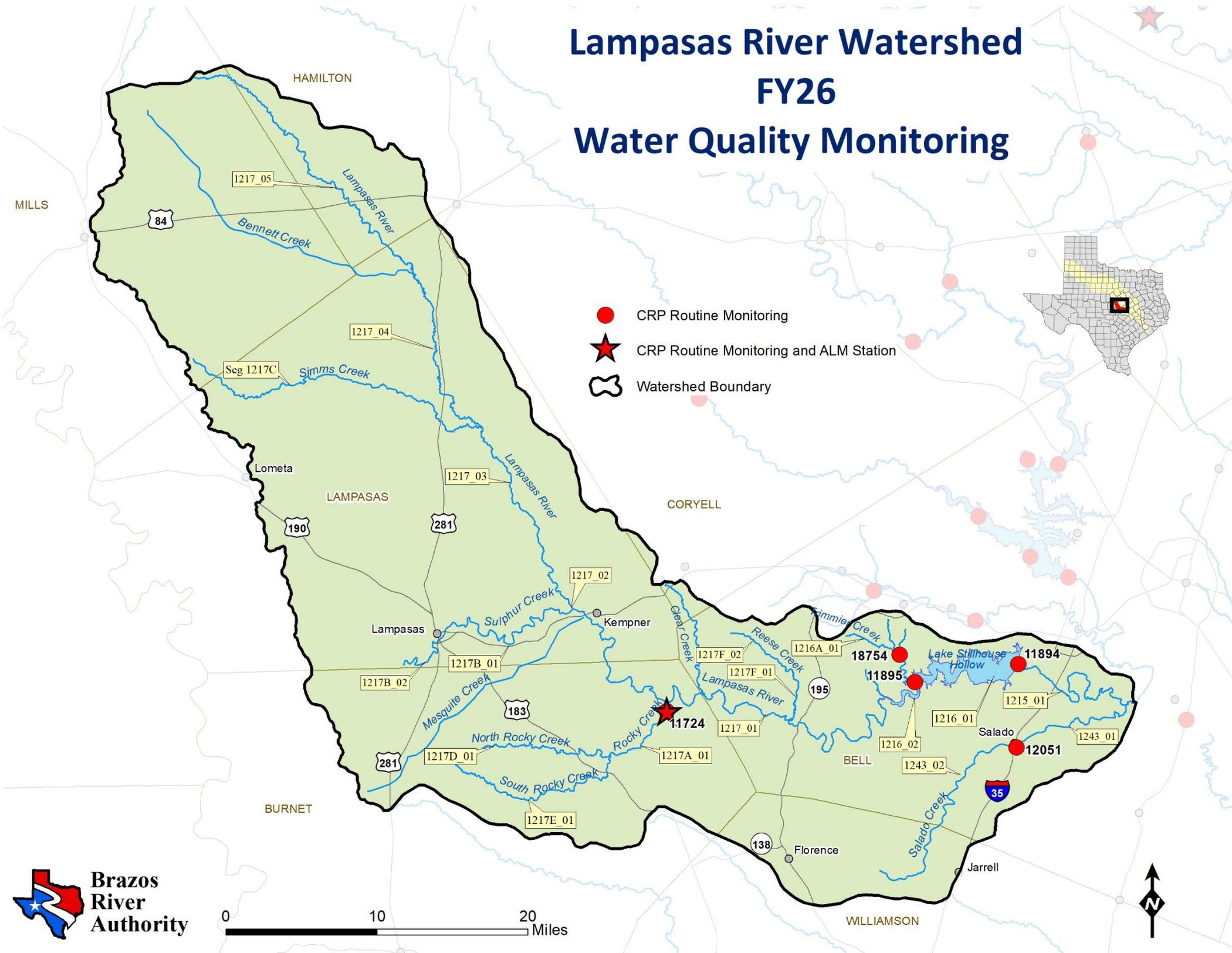
Bosque River Watershed FY26 Water Quality Monitoring



- CRP Routine Monitoring
- ★ CRP Routine Monitoring and ALM Station
- Watershed Boundary



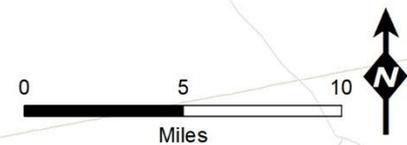
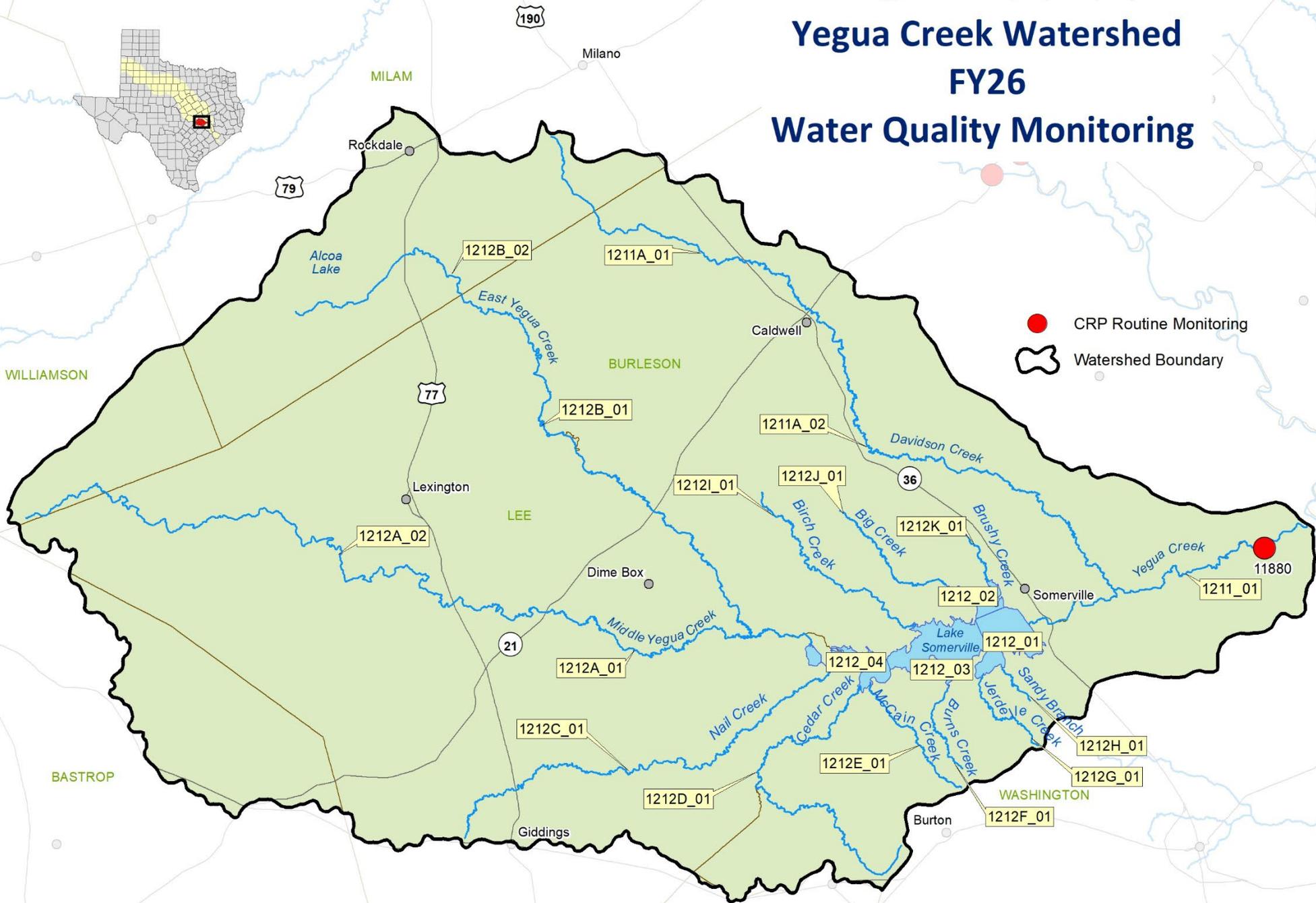
Lamparas River Watershed FY26 Water Quality Monitoring



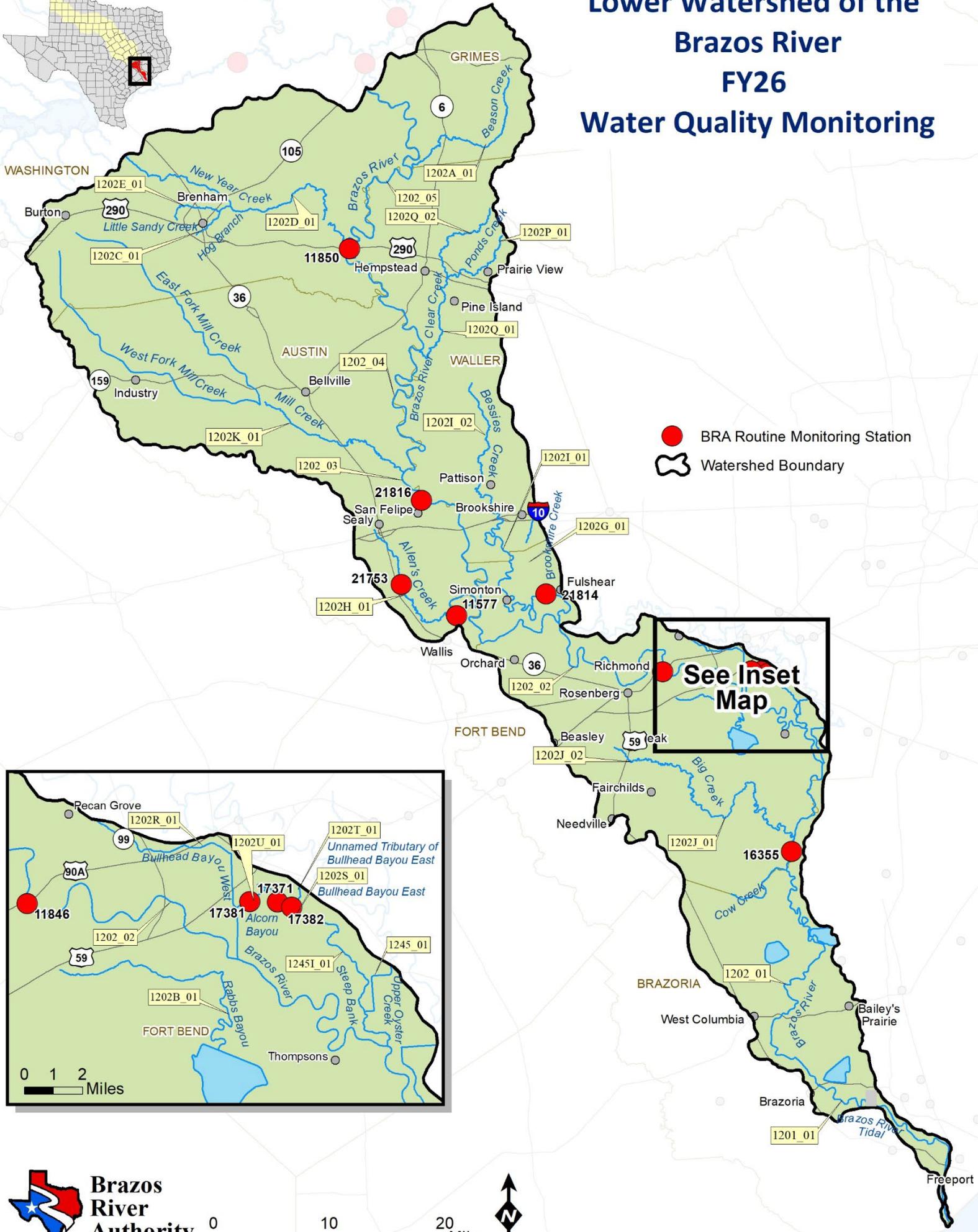
- CRP Routine Monitoring
- ★ CRP Routine Monitoring and ALM Station
- ⬭ Watershed Boundary



Yegua Creek Watershed FY26 Water Quality Monitoring

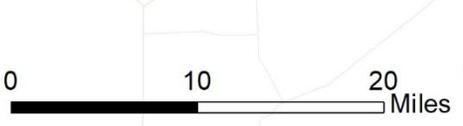
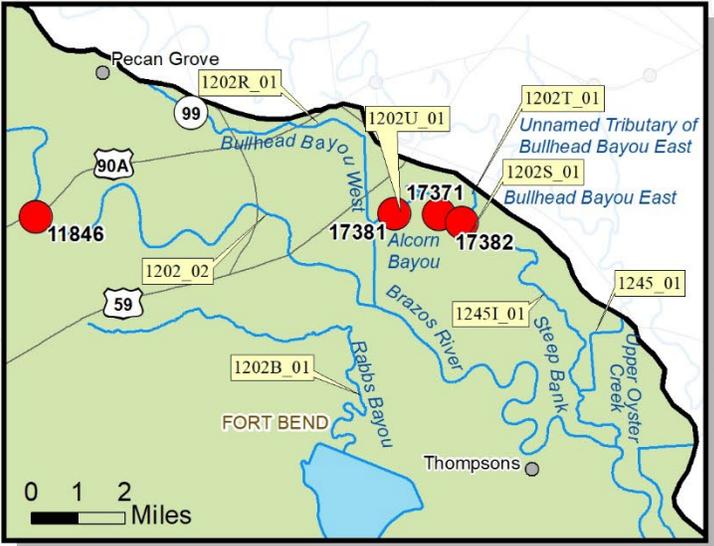


Lower Watershed of the Brazos River FY26 Water Quality Monitoring



- BRA Routine Monitoring Station
- Watershed Boundary

See Inset Map



Appendix E: Chain of Custody Forms

See Appendix D.

Appendix F: Data Review Checklist and Summary Shells

Data Review Checklist

This checklist is to be used by the Planning Agency and other entities handling the monitoring data in order to review data before submitting to the TCEQ. This table may not contain all of the data review tasks being conducted.

Data Format and Structure	Y, N, or N/A
Are there any duplicate Tag Id numbers in the Events file?	
Do the Tag prefixes correctly represent the entity providing the data?	
Have any Tag Id numbers been used in previous data submissions?	
Are Tag IDs associated with a valid SLOC?	
Are sampling Dates in the correct format, MM/DD/YYYY with leading zeros?	
Are sampling Times based on the 24 hr clock (e.g. 09:04) with leading zeros?	
Is the Comments field filled in where appropriate (e.g. unusual occurrence, sampling problems, unrepresentative of ambient water quality)?	
Are Submitting Entity, Collecting Entity, and Monitoring Type codes used correctly?	
Do sampling dates in the Results file match those in the Events file for each Tag Id?	
Are values represented by a valid parameter code with the correct units?	
Are there any duplicate parameter codes for the same Tag Id?	
Are there any invalid symbols in the Greater Than/Less Than (GT/LT) field?	
Are there any Tag Ids in the Results file that are not in the Events file or vice versa?	
Data Quality Review	Y, N, or N/A
Are "less-than" values reported at the LOQ? If no, explain in Data Summary.	
Have the outliers been verified and a "1" placed in the Verify_flg field?	
Have checks on correctness of analysis or data reasonableness been performed? e.g., Is ortho-phosphorus less than total phosphorus? Are dissolved metal concentrations less than or equal to total metals? Is the minimum 24 hour DO less than the maximum 24 hour DO? Do the values appear to be consistent with what is expected for site?	
Have at least 10% of the data in the data set been reviewed against the field and laboratory data sheets?	
Are all parameter codes in the data set listed in the QAPP?	
Are all stations in the data set listed in the QAPP?	
Documentation Review	Y, N, or N/A
Are blank results acceptable as specified in the QAPP?	
Were control charts used to determine the acceptability of lab duplicates (if applicable)?	
Was documentation of any unusual occurrences that may affect water quality included in the Event file's Comments field?	
Were there any failures in sampling methods and/or deviations from sample design requirements that resulted in unreportable data? If yes, explain in Data Summary.	
Were there any failures in field and/or laboratory measurement systems that were not resolvable and resulted in unreportable data? If yes, explain in Data Summary.	
Was the laboratory's NELAP Accreditation current for analysis conducted?	
Did participants follow the requirements of this QAPP in the collection, analysis, and reporting of data?	

Data Summary

Data Source: Brazos River Authority – LabVantage LIMS Database

Date Submitted: _____

Tag ID Ranges: _____

Date Range: _____

- I certify that all data in this data set meets the requirements specified in Texas Water Code Chapter 5, Subchapter R (TWC §5.801 et seq) and Title 30 Texas Administrative Code Chapter 25, Subchapters A & B.
- This data set has been reviewed using the criteria in the Data Review Checklist.

Brazos River Authority Data Manager: _____ **Date:** _____

Please explain in the table below any data discrepancies discovered during data review including:

- o Inconsistencies with LOQs
- o Failures in sampling methods and/or laboratory procedures that resulted in data that could not be reported to the TCEQ (indicate items for which the Corrective Action Process has been initiated and send *Corrective Action Status Report* with the applicable Progress Report).

This dataset contains data from FY2026-FY2027 QAPP Submitting Entity code BR and collecting entity BR. This is field and lab data that was collected by the Brazos River Authority. Analyses were performed by the BRA Environmental Service Laboratory unless otherwise noted. The following tables explain discrepancies or missing data as well as calculated data loss.

o **Discrepancies or missing data:**

Test	Parameter	Data Points Expected	Additional Data Points	Stations Dry, Pooled, or inaccessible (no data)	Data Points Rejected or not recorded	Data Points Submitted	Percent Data Loss* for this Dataset
Water Temperature	00010						
Reservoir Stage	00052						
Reservoir Per Cent Full	00053						
Flow	00061						
Transparency	00078						
Specific conductance	00094						
Temperature, 24HR AVG	00209						
Temperature, 24HR Max	00210						
Temperature, 24HR Min	00211						
Specific Conductance, 24HR AVG	00212						
Specific Conductance, 24HR Max	00213						
Specific Conductance, 24HR Min	00214						
pH 24HR Max	00215						
pH 24HR Min	00216						
Temperature #measurements in 24HR	00221						
Specific Conductance #measurements in 24HR	00222						

Test	Parameter	Data Points Expected	Additional Data Points	Stations Dry, Pooled, or inaccessible (no data)	Data Points Rejected or not recorded	Data Points Submitted	Percent Data Loss* for this Dataset
pH #measurements in 24HR	00223						
DO	00300						
pH	00400						
TSS	00530						
Ammonia nitrogen	00610						
Nitrate nitrogen	00620						
TKN	00625						
TP	00665						
Orthophosphate P	00671						
Chloride	00940						
Sulfate	00945						
Flow Severity	01351						
<i>E. coli</i>	31699						
Enterococcus	31701						
TDS, calc from sp. cond.	70294						
TDS	70300						
Chl <i>a</i>	70953						
Days Since Precip	72053						
Turbidity	82079						
Flow measurement method	89835						
DO 24HR Min	89855						
DO 24HR Max	89856						
DO 24HR AVG	89857						
DO #measurements in 24HR	89858						
Average Stream Width	89861						
Macrophyte Bed at Collection Point (%)	89926						
Wind Intensity	89965						
Present Weather	89966						
Water Surface	89968						

* Percent Loss = # Data Points Lost (rejected or not recorded) / # Data Points Expected for that parameter in the data set * 100%.

Discrepancy explanations

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3	
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Discrepancies or missing data for the listed tag ID

	Tag ID	Station ID	Date	Parameters	Type of Problem	Comment/PreCAPs/CAPs
5						
6						